# Strange tadpoles from the lower Miocene of Turkey: Is paedogenesis possible in anurans?

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Fossil material from the lower Miocene collected in the basin lake of Beşkonak (Turkey) included 19 slabs showing 19 amphibian anuran tadpoles of rather large size, at Gosner stages 36–38. These well preserved specimens show many morphological and skeletal characters. They are here tentatively referred to the genus *Pelobates*. Two of these tadpoles show an unusual group of black roundish spots in the abdominal region, and a third similar group of spots is present in another slab but we were unable to state if it was associated with a tadpole or not. Several hypotheses can be proposed to account for these structures: artefacts; intestinal content (seeds; inert, bacterial or fungal aggregations; eggs); internal or external parasites; diseases; eggs produced by the tadpole. The latter hypothesis is discussed in detail and is shown to be unlikely for several reasons. However, in the improbable case where these spots would correspond to eggs, this would be the first reported case of natural paedogenesis in anurans, a phenomenon which has been so far considered impossible mostly for anatomical reasons (e.g., absence of space in the abdominal cavity).

Key words: Amphibia, Anura, egg, fossil, paedogenesis, tadpole, Miocene, Turkey.

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## Introduction

Because they are composed of about 90% water (Bradford 1984), tadpoles of anuran amphibians are extremely fragile organisms: just a few hours after having been taken out of water and deposited on the ground, virtually nothing remains of them. Therefore tadpoles, although periodically very abundant in some continental aquatic habitats, are not good candidates for fossilisation, and only relatively few fossil tadpoles are known, presumably having benefited from unusual fossilisation conditions. Among the best preserved ones are the lower middle Miocene fossil tadpoles from two localities of the Gürcü Valley (40°35–38'N, 32°39–42'E, altitude 1200–1300 m), about 90 km north of Ankara, Turkey, described and illustrated by Wassersug and Wake (1995), and those from a nearby locality, the basin lake of Beşkonak (40°38'N, 32°41'E), also situated in the Gürcü Valley, reported by Rückert-Ülkümen et al. (2002). In both cases, the tadpoles were referred to the genus *Pelobates* Wagler, 1830 (Pelobatidae Bonaparte, 1850). The age of the fossil deposits of the Gürcü Valley was estimated to 14.1 ± 1 Mya, corresponding to the middle Miocene, dated using K/Ar isotopes of biotites contained in the rocks (Kasapligil 1981), but this date was challenged by subsequent studies of the flora and of the insect fauna (Paicheler 1988; Nel and Paicheler 1993), which suggest that these deposits are from the Oligocene–Miocene boundary or the lower Miocene.

We here report on several well preserved anuran tadpoles also from the basin lake of Beşkonak (Fig. 1). These specimens were collected by one of us (JCP) in 1972, but never described in detail, although they were briefly mentioned in a general paper dealing with the fossils of this locality (Paicheler et al. 1978). The fossilisation occurred as a result of volcanic activity which poured lava and hot volcanic ash into the lake, causing rapid sedimentation of diatomaceous frustules. These, along with the accumulation of fine silt from streams flowing into the lake, formed the laminated diatomites and paper shales (Kasapligil 1981). The excellent preservation of the animals also suggests reducing conditions on the bottom of the lake with a $H_2S$ layer (Paicheler et al. 1978).

The fauna and flora found with these tadpoles are all aquatic organisms or organisms linked to aquatic habitats. This area was therefore presumably flooded at the time of fossilisation. According to the floristic, palynologic and entomological analyses, the regional climate was hot temperate of a Mediterranean type (Paicheler et al. 2007). This area was subject to regular alternation of dry and wet seasons. Quite shallow, the lake presumably varied from a great lake connected with other basins to a huge swamp, depending on the season (Paicheler et al. 1978).
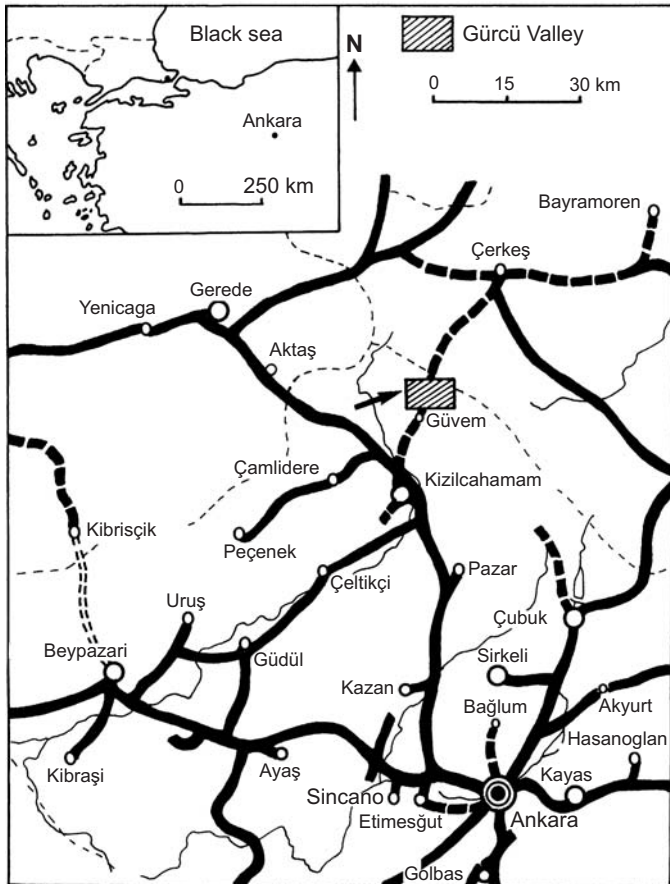
Fig. 1. Map of Northwestern Turkey showing the area which yielded the fossil tadpoles in this study, near Güvem, in the Gürcü Valley. See Fig. 2 for detailed map of this area.

The tadpoles appear as organic film imprints within a diatomite matrix. They are well preserved, lying on their side, on their venter or ventrolaterally and thus showing details of their soft anatomy, such as gills, eyes, lungs, parts of the intestinal tract and blood vessels, caudal myotomes and caudal fins. Bones and cartilage present some depressions. The slabs that contain the tadpoles also yield plant remains and a tiny bird feather, but no adult amphibians. However the most striking feature of these specimens is that two of them show the clear presence of rounded spots in the abdominal region. Though there are several alternate explanations, the question arises whether these spots can be interpreted as eggs produced by the tadpoles.

*Abbreviations*.—BL, body length; MNHN, Muséum National d'Histoire Naturelle, Paris, France; UCMP, University of California Museum of Paleontology, Berkeley, USA.

# Material and methods

We examined traces of fossil tadpoles from 18 distinct slabs over several thousand slabs containing tadpoles observed in the field in this region in 1972 by JCP. These slabs with tad-poles were collected in two different localities of this basin: Ahlath Dere, north of the village of Kerimler; and west of the village of Agaöz (Fig. 2). Both localities are from the same stratigraphic level (Paicheler 1988). We identified 19 individual tadpoles in these slabs. These slabs, as well as another one showing a group of roundish spots, discussed below, are housed in the Paris Museum collection (Reptiles and Amphibiens), under the numbers MNHN 1998.9173–9194.

The tadpoles were observed under a binocular Leica MS5 and measured with a hand calliper due to their large size. Based on their cranial ossification, they were tentatively referred to the standard developmental stages of the table of Gosner (1960).

# Description of the material

**The tadpoles**.—The 19 tadpoles are well preserved and do not show distortions or crushing. They are similar in aspect, without any trace of posterior limb buds, thus suggesting that they were in rather early stages of larval development, before the beginning of the period of metamorphosis. However, they show a somewhat extended range of sizes, so that it can be hypothesized that they were not all of the same age, and
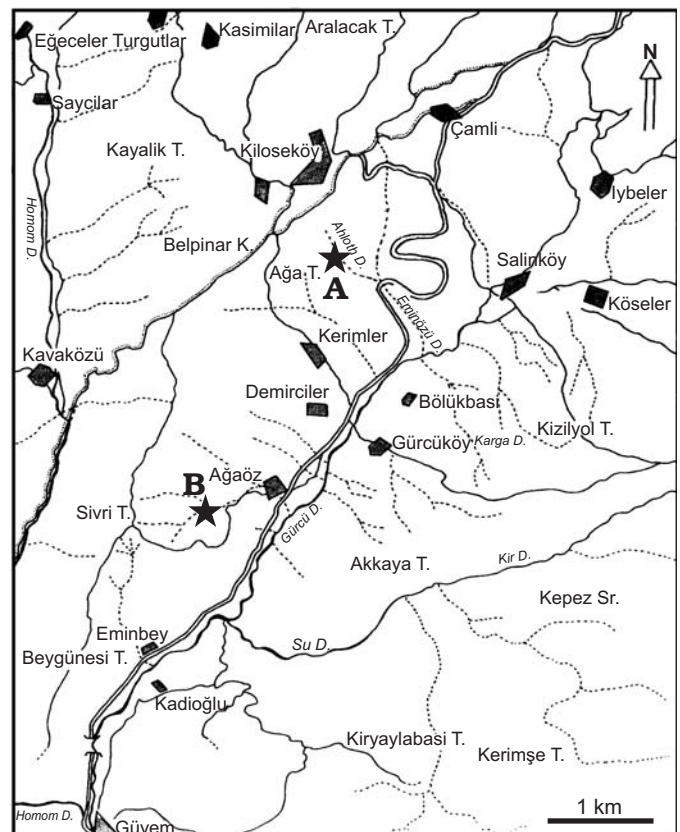


Fig. 2. Sites of collection of the fossil tadpoles in the Gürcü Valley studied here (enlarged from Fig. 1), Turkey. **A**. Ahlath Dere, where the specimen MNHN 1998.9173 was collected. **B**. Agaöz, where the specimen MNHN 1998.9174 and the slab with the mass of roundish spots MNHN 1998.9191 were collected.

therefore originated from different egg clutches, laid at different dates. The absence of hindlimb traces precludes their assignation to an accurate developmental stage. However, some indices could help to determine a range of developmental stages (see below).

Anteriorly, the keratinised beaks are visible in a few specimens, as well as the serrations in one of them. The upper beak seems to have the form of a large arch and the lower beak of a broad "U". Keratodonts have not been observed.

Skeletal elements are visible in almost all tadpoles, especially the frontoparietal and the vertebrae, although the parasphenoid and the otic capsules are also visible in a few. The frontoparietal bones lie laterally to the cultriform process of the parasphenoid; they are elongate, drop-shaped, tapering anteriorly and enlarged posteriorly to a blunt extremity. The lateral notches present on the frontoparietal of the specimen UCMP 139183, described in detail by Wassersug and Wake (1995), are absent in all of our specimens exhibiting frontoparietal bones. However, these specimens exhibit a small concavity which could correspond to the fontanelle. The parasphenoid is well visible in one specimen. It has the form of an inverted T, with the cultriform process long and fine and ending in a point anteriorly. The alar processes are fine triangles with a pointed tip. The posterior end of the parasphenoid forms an obtuse angle.

Imprints of otic capsules are present in some fossils. They are rounded structures situated between the frontoparietals and the alar processes of the parasphenoid and lateral to them. They are always smaller than the eyes, from about half size to almost similar size.

Between eight and eleven vertebrae are observable in these specimens. The first vertebra, the atlas, is not well outlined but seems thicker than the following ones, although what we observed could be a print of the atlas and of the first post-atlas vertebra. Each vertebra bears well developed transverse processes, the diapophyses. The diapophyses of the three first vertebrae are straight, the following narrowing in their middle, then widening in their distal part. Furthermore, their length decreases in an anteroposterior direction along the vertebral series although the size of all vertebrae is equal. The last vertebra is smaller than the others in one specimen. Along the vertebra series, the imprint of the notochord is often visible.

The absence of hindlimb traces suggests that these tadpoles are in a rather early developmental stage, at least before stage 30 in which the hindlimb buds are rather developed (Gosner 1960). However, the ossification of skeletal elements described above gives some contradictory indication upon their developmental stage. In several families of living anurans, the first three osseous structures that appear during the ossification of the skull are the otic capsules, the parasphenoid and the frontoparietals, which develop during the stages 36–38 and are present at the latest at the end of stage 38 (Kemp and Hoyt 1969; Gaudin 1973, 1978; Hanken and Hall 1984, 1988; de Sá 1988; Wiens 1989; Trueb and Hanken 1993; Maglia and Púgener 1998). We therefore conclude that the Turkish tadpoles are in developmental stages 36–38.

The same interpretation was proposed by Wassersug and Wake (1995) for their tadpoles, although in this case also no hindlimb structures were visible. In fact, we know of no instance of fossil tadpoles at these stages showing hindlimbs: among the many photographs of tertiary tadpoles shown in the book by Špinar (1972), the only ones that show hindlimbs are at later stages, just before the end of metamorphosis.

**The abdominal roundish spots**.—Among the 19 tadpoles examined, two show a very striking peculiarity, i.e., the presence of roundish dark spots in the abdominal region.

The first specimen (MNHN 1998.9173, BL 37.2 mm), collected in locality (A) Ahlath Dere (Fig. 2), shows about 100 such spots concentrated in the posteroventral part of its abdomen, a part of the tadpole's body which is usually entirely filled up with the intestine (Fig. 3). These spots are all of similar aspect and size, very regular in shape, roundish and with a diameter of 0.76–1.06 mm (n = 7). They are as dark as the eye imprints of the tadpoles, and darker than the remains of other organs and soft parts, suggesting either a protein-rich structure or more probably that they are due to a bacterially induced pyritization (Briggs 2003). Some of these spots appear isolated, whereas others appear in close contact or even connected to each other. The abdomen of this specimen has a fully normal shape, not being particularly expanded or deformed by the presence of these structures.

The second specimen (MNHN 1998.9174, BL 36.6 mm), collected in locality (B) Agaöz (Fig. 2), shows only a few spots (roughly 13), with a similar shape and slightly larger (diameter 0.91–1.37 mm; n = 4) than in the first specimen (Fig. 4). These spots are located ventrally and in the posterodorsal part of the abdomen, which also has a normal shape.

Finally, on a slab (MNHN 1998.9191) also collected in Agaöz, a third group of similar spots is present but as all the surrounding (and a part of this group of spots) is missing, we are unable to state if it was also present within a tadpole or not (Fig. 5). The number of spots is estimated to more than 150 and their mean diameter is 1.3 mm (1.0–1.6 mm; n = 8). The aspect and size of these spots suggest that they are of the same nature as those observed in the two tadpoles.

Unfortunately, the preservation of these soft structures as a film rather than as three dimensional structures prevents us from extracting more information from these spots than the observation of these structures, and hypotheses have to be proposed to explain their presence.

Whatever the origin of these spots is, it is worth noting that none of the 218 slabs bearing fossil actinopterygian Cyprinidae Cuvier, 1816 (all referred to *Leuciscus* Cuvier, 1816) collected with these fossil tadpoles, presents such structures (Gaël Clément, personal communication 2008).

# Discussion

**Identification of the tadpoles**.—Taxonomic identification of these tadpoles is difficult. No specimen shows well-pre-
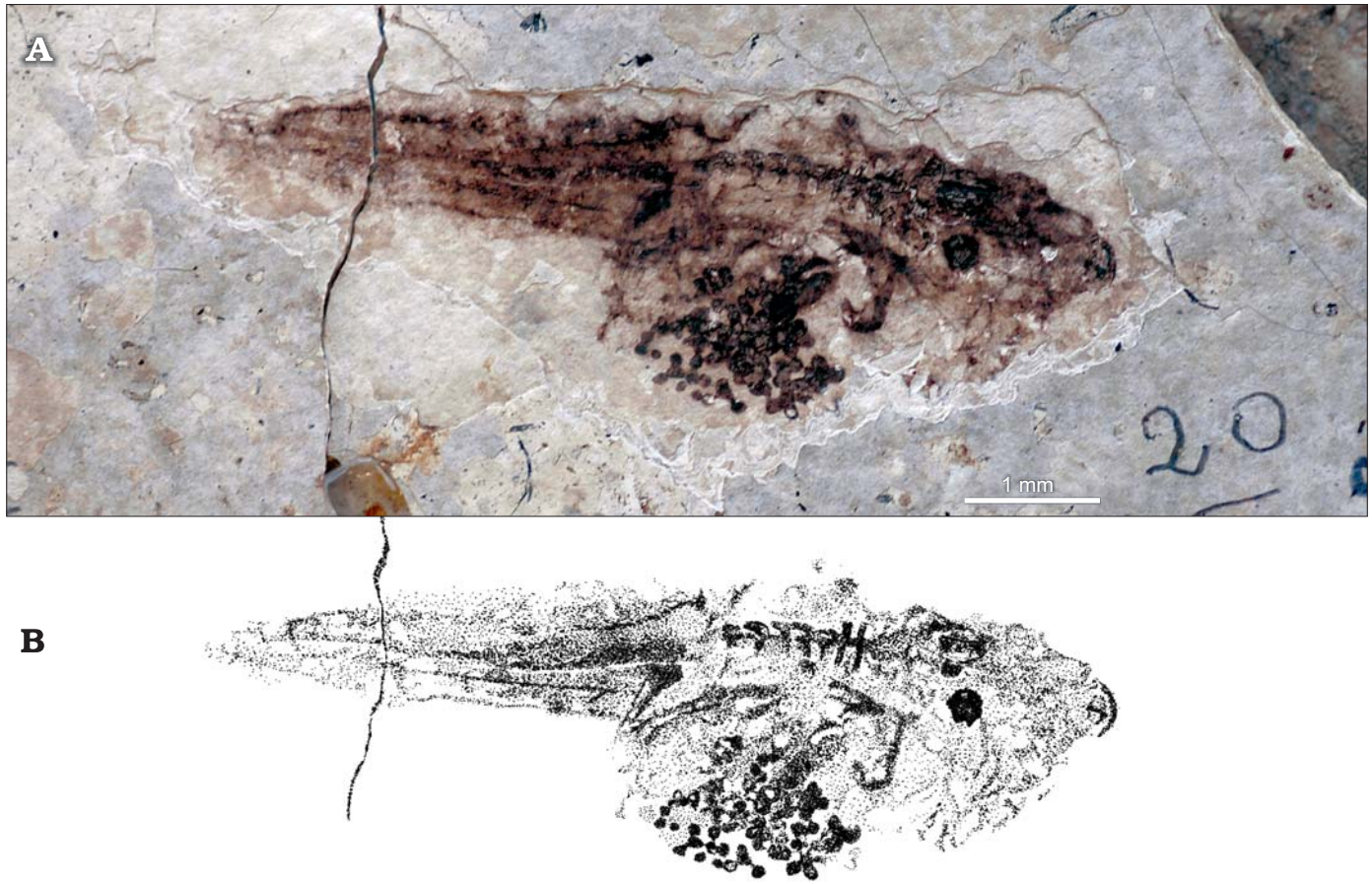
Fig. 3. Photograph (**A**) and drawing (**B**) of the fossil tadpole MNHN 1998.9173 from Ahlath Dere, Gürcü Valley, lower Miocene, Turkey, bearing a group of spots in the abdominal region.

served mouthparts with keratodont rows that would allow their straightforward allocation to a genus. The large size of some of these tadpoles (about 90 mm; see Table 1) excludes a number of extant anuran genera, but leaves several possibilities. Tadpoles of the genera *Alytes* Wagler, 1829 (Alytidae Fitzinger, 1843), *Pelobates* (Pelobatidae), *Pelodytes* Bonaparte, 1838 (Pelodytidae Bonaparte, 1850) and *Pelophylax* Fitzinger, 1843 (Ranidae Rafinesque-Schmaltz, 1814) are known to reach similar large sizes: *Alytes obstetricans* (Laurenti, 1768), 90 mm (Boulenger 1891); *Pelobates fuscus* (Laurenti, 1768), 200 mm (Grillitsch et al. 1983); *Pelobates syriacus* Boettger, 1889, 205 mm (Boschwitz 1957); *Pelodytes punctatus* (Daudin, 1802), 95 mm (van den Elzen

Table 1. Some measurements (in millimetres) of fossil tadpoles at Gosner (1960) stages 36–38 from the basin lake of Beşkonak. Only the 6 most complete tadpoles were measured.

| Tadpole number | Total length (TL) | Body length (BL) | Tail length (TAL) | Tail height (MTH) | Ratio BL/TAL (%) | Ratio MTH/TAL (%) |
|---|---|---|---|---|---|---|
| MNHN 1998.9173 | | 37.2 | | | | |
| MNHN 1998.9174 | 92.0 | 36.6 | 55.4 | | 66 | |
| MNHN 1998.9175 | | | 49.5 | 17.7 | | 36 |
| MNHN 1998.9176 | | 47.4 | | | | |
| MNHN 1998.9177 | | 42.2 | | | | |
| MNHN 1998.9178 | 92.7 | 35.1 | 57.6 | | 61 | |

1976); *Pelophylax esculentus* (Linnaeus, 1758), 163 mm (Borkin et al. 1982); *Pelophylax ridibundus* (Pallas, 1771), 165 mm (Borkin et al. 1982); and *Pelophylax perezi* (Seoane, 1885), 90 mm (Llorente et al. 1995). All these tadpoles are usually smaller than the sizes just listed, but can reach them when they overwinter without metamorphosing (Boulenger 1891; van den Elzen 1976; Savage 1952; Grossenbacher 1997; Kuzmin 1997; Miaud and Muratet 2004). In normal conditions however, the species of the genus *Pelobates* are those which reach the largest size before metamorphosis among western Palearctic frog tadpoles (Díaz-Paniagua 1988). Tadpoles without a thyroid gland or with a deficiency in the hormonal system (axis hypothalamus-hypophysis-thyroid) are known to reach sometimes unusual large sizes, e.g., four times longer than in normal tadpoles in *Xenopus laevis* (Daudin, 1802) (Boschwitz 1957; Borkin et al. 1982; Rot-Nikcevic and Wassersug 2004), but this does not seem to be the case in our fossil tadpoles (see Suggestions below).

Palaeontological and distributional data also allow some inferences. The continental Miocene of Turkey is marked by major faunal interchanges between Africa and Laurasia, so the fossil tadpoles can also have a north African origin, and the anuran genera present in this region are considered here as well as the native fauna from Turkey. The genus *Alytes* can rather safely be excluded, as it is currently know only
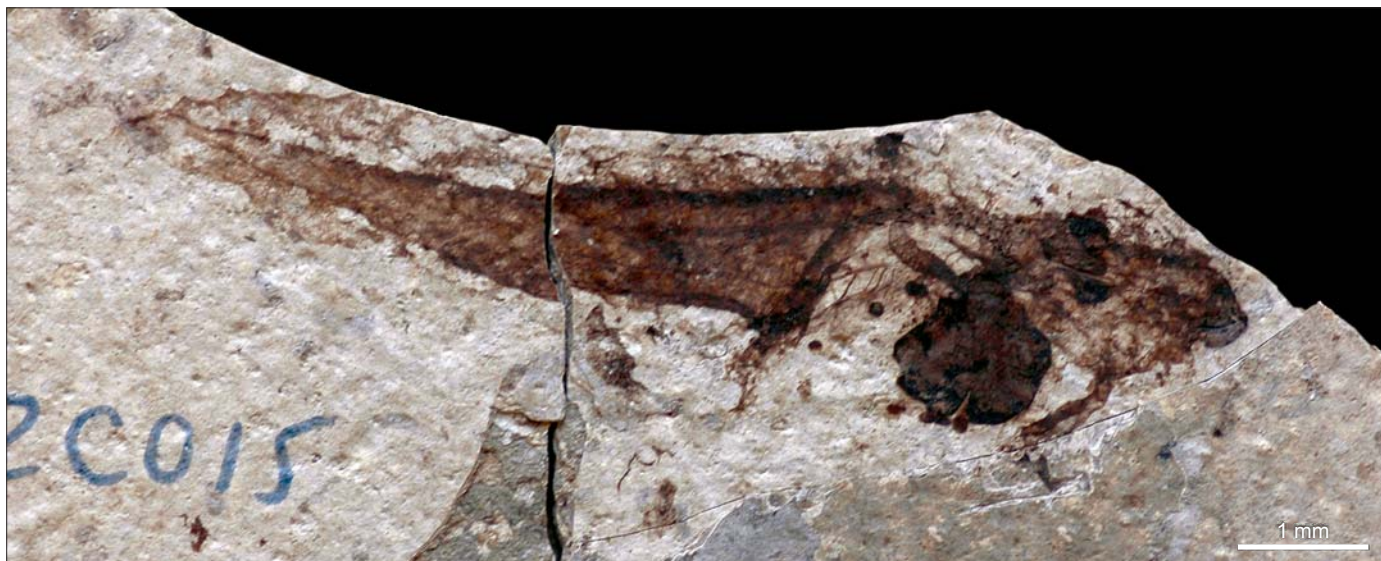
Fig. 4. Photograph of the fossil tadpole MNHN 1998.9174 from Agaöz, Gürcü Valley, lower Miocene, Turkey, bearing a few spots in the abdominal region.
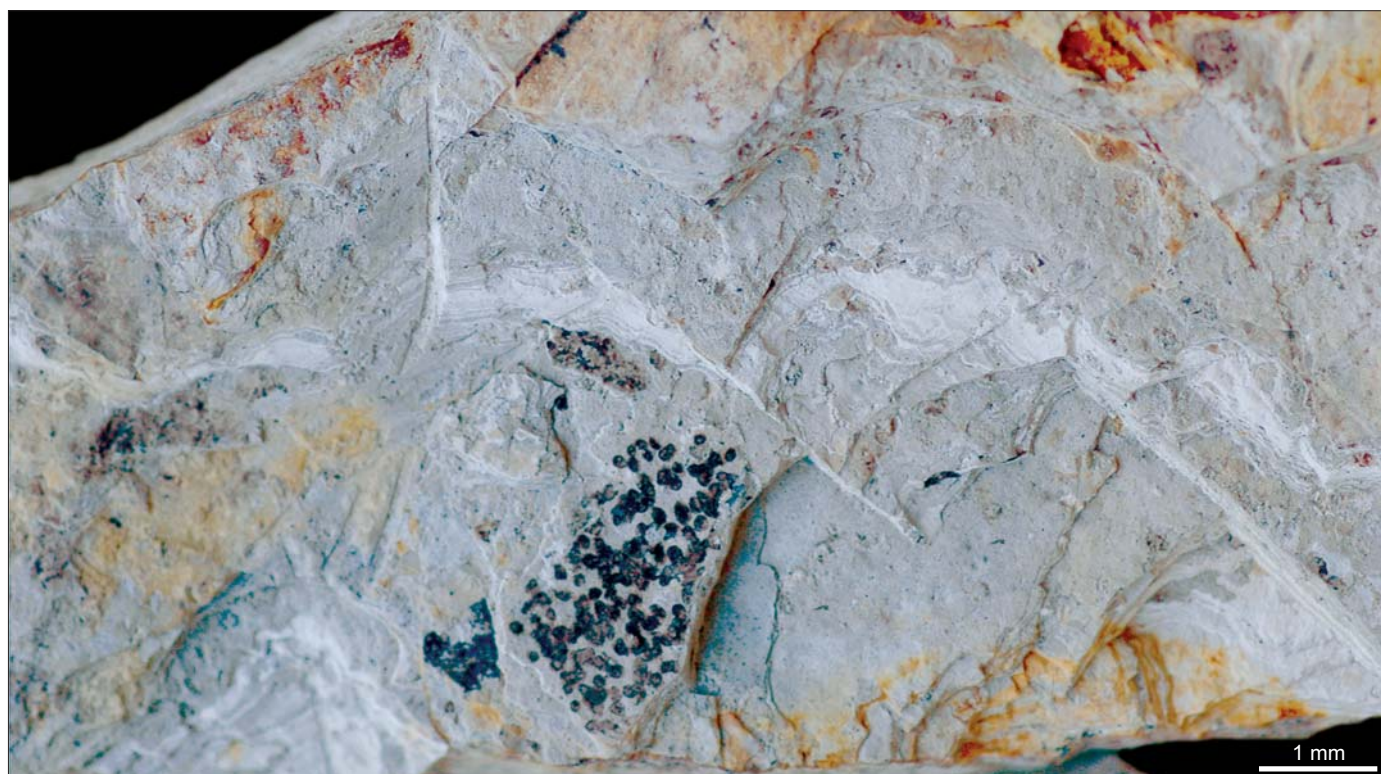


Fig. 5. Photograph of the isolated group of spots (MNHN 1998.9191 from Agaöz, Gürcü Valley, lower Miocene, Turkey).

west of the Alps and in northern Africa, but no fossil of this genus is known from the eastern Mediterranean region. The other three groups are currently represented in the native fauna of Turkey (Basoglu and Özeti 1973; Baran and Atatür 1998), so that they can remain potential candidates. Fossils of the family Pelobatidae are known since the early Eocene (MP 7) for *Eopelobates* Parker, 1929 and the early–middle Oligocene (MP 22 or 23, 30–31 Mya) for *Pelobates* (Rage and Roček 2003). Fossils of Pelodytidae are known since the late Eocene (MP 16) or perhaps middle Eocene (MP 13, about 40 Mya), and since the middle Miocene (11–12 Mya) for *Pelodytes* although this genus is much older (Rage and Roček 2003). Finally, green frogs of the genus *Pelophylax* (Frost et al. 2006) are known since the early Oligocene (MP 22, 32–33 Mya) (Rage and Roček 2003).

We measured preserved tadpoles (kept in the MNHN) of all the recent species listed above and also of *Pelobates cultripes* (Cuvier, 1829), *Pelodytes caucasicus* (Boulenger,

1896) and *Pelophylax saharicus* (Boulenger, 1913) (data not shown) to compare these data with those of Table 1. The results do not allow exclusion of any of these four genera. However, the general shape of the body and of the tail in lateral view suggests that the fossil tadpoles cannot belong to the genus *Pelophylax*: in the *Pelophylax* tadpoles the height of the upper tail fin shows its maximum in its proximal part (Nöllert and Nöllert 1992: 114) whereas in the Turkish tadpoles, the maximum height of the upper tail fin is reached more posteriorly. Another frog family should be discussed here, the Palaeobatrachidae Cope, 1865, which were common in Europe during the Tertiary, although not reported from Turkey (Sanchíz 1998). Assignation of the fossil tadpoles to this extinct family can be excluded, as the tadpoles of Palaeobatrachidae exhibit several morphological and anatomical characters which easily distinguish them from those of other families, and especially the Pelobatidae. The most conspicuous are the presence of free ribs, the absence of lateral processes of the parasphenoid, an undivided frontoparietal and the general shape of the body which is reminiscent of the Pipidae Gray, 1825 (Špinar 1972: 38). Furthermore, although not clearly mentioned in the literature, the tadpoles of the Paleobatrachidae should lack keratinised mouthparts, as in all members of the superfamily Pipoidea Gray, 1825.

Wassersug and Wake (1995), reporting about similar tadpoles from a nearby locality, referred them to the genus *Pelobates* for several reasons that they explained in detail. The evidence in favour of this identification is rather convincing. We observed no character suggesting that our tadpoles would not be conspecific with those described by these authors. However, these authors did not discuss the possibility that these tadpoles belong to the extinct pelobatid genus *Eopelobates*. Maus and Wuttke (2002) compared osteological features of tadpoles of *Pelobates* and *Eopelobates* and found two characters which allow discrimination of the two forms: the shape of the frontoparietal bones, which possess a processus lateralis superior in *Pelobates* species but not in *Eopelobates*, and the width/length ratio of the vertebrae II–IV which is between four and five in *Pelobates* and between five and seven in *Eopelobates*. The specimen MNHN 1998.9176 in which the frontoparietals are well preserved shows a processus lateralis superior on the right one (the tadpole being slightly in latero-dorsal view, this processus is hidden in the left one). The vertebrae II and III of the same specimen, although their length is not very well outlined, have a width/length ratio slightly below five.

Since all these data do not contradict the hypothesis of Wassersug and Wake (1995), we propose to adopt it for our tadpoles also, pending its possible refutation. Rückert-Ülkümen et al. (2002) reached the same conclusion for similar tadpoles from a nearby locality. Furthermore, as stated above, our sample is composed of tadpoles encompassing a large size range but seemingly of about the same developmental stage. The presence of tadpoles of different sizes in a waterbody seems to be typical for *Pelobates fuscus*, which is an additional argument for the allocation of the fossil tadpoles to the genus *Pelobates* (Frank Glaw, personal communication 2009).

**Hypotheses about the spots**.—Several hypotheses can be put forward to try and explain these spots. Let us first note that these were observed in two distinct localities which are more than 2 km apart (Fig. 2), so that the cause responsible for their appearance cannot be taken as exceptional, e.g., linked to a particular event or to closely related tadpoles (i.e., produced by a single female).

*Artefacts*.—The dark spots could be non-organic artefacts produced during the fossilisation process, such as gas vesicles originating from decomposition of the intestinal contents of the tadpoles. However, we do not know of any other occurrence of such spots and we are not aware of a natural process that could generate such regular spots. Furthermore, the dark aspect of these spots suggests a protein-rich structure, which does not fit with this hypothesis.

*Intestinal contents*.—The location of the spots within an area of the tadpole's body (lower posterior part of abdomen) that in normal tadpoles is entirely occupied by the coiled intestine, suggests that the spots could correspond to structures that were part of the intestinal contents. Usually intestinal contents of beaked anuran tadpoles are composed of pieces not exceeding 6 mm in diameter (Heyer 1973: fig. 8), but only few works have been devoted to the detailed study of the nature of tadpoles' intestinal contents. Among the other tadpoles found in Gürcü, intestinal remains are visible in some specimens (MNHN 1998.9176, 9179) as loose filamentous structures, very different from the dark spots here discussed. A striking feature of these dark spots is their regularity in shape, but with a rather high size variability (diameter from 0.76 to 1.6 mm). Such spots could possibly be viewed as a mass of seeds that would have been swallowed and kept unaffected by the whole digestion process. However, the spots appear darker than all vegetal remains present in the slabs, suggesting richer protein content than usually present in vegetal material. Estes et al. (1978) reported on small hollow spherical bodies lateral to the right posterior region of the vertebral column of a *Shomronella jordanica* Estes, Špinar, and Nevo, 1978 (Pipidae) tadpole in dorsal view. These authors interpreted these structures as part of the intestinal content. Independent of their arguments to reach this conclusion, these structures are quite different from those observed in our samples in that they are clearly hollow in the *Shomronella* Estes, Špinar, and Nevo, 1978 tadpole whereas the roundish spots discussed here are uniform. Other possibilities for intestinal contents would include either inert structures or bacterial, fungal or algal aggregations developed on intestinal vegetal debris, or eggs.

*Internal or external parasites*.—We are not aware of any published observation of parasitic infection of tadpoles that would produce a mass of numerous parasitic cysts of regular shape that would correspond to those observed in our two specimens. As for the possible hypothesis of external parasites that would have been fixed to the skin of the belly of the tadpoles, it would not correspond to any known observation in the living fauna. This is quite unlikely given the very thin skin of this

part of the tadpole's body, which presumably would not long resist the fixation of external parasites without leading to a tearing of this skin and to the death of the tadpoles.

*Disease*.—The spots could be thought of as pathological nodules, such as tumours. We do not know of any known case of multiple nodules of similar and regular aspect in tadpoles.

*Eggs*.—The hypothesis that seems to best fit the observations is that the dark spots correspond to eggs that would have grown in the abdomens of the two tadpoles MNHN 1998.9173–9174. This would explain the large number of spots of similar aspect, with a roundish shape. Eggs would be composed mostly of vitellus, a protein-rich material likely to produce a dark fossil trace. The two groups of eggs associated with tadpoles would still be located in the ovaries, whereas the third group not clearly located within a tadpole might possibly represent a clutch of eggs after its laying by another tadpole. Although less unlikely than the other hypotheses that we have considered above, this hypothesis still raises a number of questions, that will now be considered in more detail.

**A review of fossil lower vertebrate eggs**.—In his review of fossil vertebrate eggs, Hirsch (1994) did not mention anamniote eggs. However, a few reports of anamniote aquatic vertebrate fossil eggs exist in the literature.

Grande (1984: 101, fig. 2.46) reported the discovery of a specimen of, probably, *Diplomystus dentatus* Cope, 1877, a clupeomorph teleost, fossilised inside the eggs, from the early Eocene of Wyoming (USA). He provided a photograph of a small fossil teleost curled up inside a light coloured spot. The "eggs" are 12 mm in longest diameter. The fish does not appear to be inside an egg because of its size (about 18 mm) and its degree of ossification (the caudal fin and pectoral girdle are present). Clupeomorph eggs are tiny, never exceeding 2 mm in diameter in extant taxa. We suggest the spot has another origin.

Gall (1991: fig. 2) found "fish eggs" in the Grès à *Voltzia* Formation from the lower Triassic of France. The extremely well preserved fossils show eggs, probably of vertebrates.

Martill (1993) described fossil eggs from the Early Cretaceous Santana Formation of Brazil. He provided SEM micrographs of what undoubtedly are three-dimensional preserved eggs probably produced by an actinopterygian.

Mamay (1994) described Pennsylvanian fossil eggs from New Mexico (USA) of probable chondrostean origin that he considered "the oldest known vertebrate egg masses".

Clément (1999) provided an explanation of structures sometimes found in the abdominal region of fishes (sarcopterygians as well as actinopterygians). These structures appear as rounded, regular traces. After finding these structures in a fossil coelacanth, he considered the possibility of them being eggs, but he discarded this explanation for several reasons, and favoured a gas origin for the structures. This interpretation explains why traces are often carved in bones.

Finally, three reports of fossil eggs assigned to amphibians are known to us.

Godfrey (1992, 1995) described fossil eggs from the Pennsylvanian of Illinois (USA) that he considered as possible amphibian eggs from now extinct groups.

Mamay et al. (1998) described eggs from the Lower Permian of Texas (USA) that he assigned to the dissophoroid temnospondyl amphibians, also an extinct taxon.

The last reference is the only one that concerns fossil anuran eggs. Špinar (1972) provided photographs of what he thought to be eggs, placed at both sides of the abdominal cavity of an adult anuran from the Tertiary of central Europe. However, both their size and the fact that they appear as carved traces in bones identical to those known in fishes suggest that the same phenomenon implied by Clément (1999), i.e., gas vesicles, was responsible for these traces.

Thus, none of the observations yet published bears a close resemblance with our observations on the Gürcü Valley tadpoles.

**Critical study of the egg hypothesis**.—First of all, it should be stressed that if the dark spots observed here really represented eggs in ovaries of tadpoles, this would be an extraordinary discovery. In amphibians, this phenomenon is well known in urodelans, but very rare in anurans. Maturation of gonads containing mature gametes have been observed in giant, non metamorphosing specimens of laboratory breeding colonies of *Xenopus leavis* (Rot-Nikcevic and Wassersug 2004) and in natural populations of the European water frogs of the *Pelophylax esculentus* complex (Borkin et al. 1982).

The ability for an organism to reproduce while being still in a larval condition (in regards to somatic features) is part of the vast array of developmental phenomena under the general term of *aneuchrony* (Dubois 1987), i.e., in which chronology of development is modified compared to ancestral forms. This modification may have two different aspects: (1) *acceleration* or *retardation* of development relative to ancestors; (2) presence or absence of *dissociation* between traits (e.g., somatic vs. gonadal) during chronology of development relative to ancestors. Aneuchrony is generally referred to as *heterochrony* (e.g., Gould 1977; Pierce and Smith 1979; Alberch et al. 1979), but the latter term should better be used to refer to dissociation between traits during the chronology of ontogeny, so that *homochronic aneuchrony* (without dissociation between traits) can be distinguished from *heterochronic aneuchrony* (with dissociation between traits) (Dubois 1987).

Paedogenesis (as defined by Dubois 1987) is just one among several possible types of dissociation between traits during ontogeny, in which sexual maturity (mature gonad), with emission of eggs liable to develop, is attained by an organism still showing an overall larval phenotype. This is a rather common phenomenon in urodelan amphibians, but it has never been described in anurans. In the latter group, only a limited number of cases of "partial paedomorphism" (Dubois 1987) have yet been reported, including adult anurans still showing a tail (Olivier 1893), or with a forelimb still emprisoned under skin (Richardson and Barwick 1957; Dubois 1979), or with an intestine still coiled like in tadpoles (Breuil

1981). Similar phenomena (e.g., retention of tail after metamorphosis) were also observed in transgenic tadpoles (Huang et al. 1999; Huang and Brown 2000), or induced by chemical treatment (Elinson et al. 1999), or found in rare athyroid tadpoles appearing spontaneously in laboratory colonies (Rot-Nikcevic and Wassersug 2004). The latter died at the stage when the tail would normally be resorbed.

All these are cases of isolated larval somatic characters retained in an otherwise adult organism, but no cases are known in anurans of the reverse situation, i.e., reproduction in larval organisms. Wassersug (1974, 1975) suggested that this was impossible in anuran tadpoles for anatomical reasons, the abdomen being completely filled up with the intestine, which would leave no space for the development of the gonads. One purely theoretical possibility would be to provide space for the development of the gonads by ceasing to feed, as tadpoles kept without any food tend to show a depletion of the abdomen (before finally dying). The fact that some tadpoles, under special conditions, proved able to develop mature gonads, although without ability to reproduce, raises other questions.

If this phenomenon was possible, why has it never been observed in nature in tadpoles of living anurans species? Such a possibility, if it existed, would have been likely to have been "exploited" in the course of the evolution of anurans, as it has been in that of the urodelans. In the latter group, paedogenesis occurs in some taxa subject to very severe environmental constraints in the aerial stage of their cycle, whereas the aquatic environment is rich and stable. As summarised by Gould (1977), for example, in such conditions, slow growing individuals keep their larval habitat rather than trading it for a hazardous environment. This adaptive response proved so efficient in urodelans that it affected not only some populations and species, but became fixed in more important clades: some families of this order (Proteidae Gray, 1825 and Sirenidae Gray, 1825) are considered to be paedogenetic in origin (Duellman and Trueb 1985; Frost et al. 2006). The fact that not a single case of paedogenesis (heterochrony with possibility of reproduction) is known in anurans suggests that it is unlikely that the Gürcü tadpoles, even if they developed eggs in their abdomen, were able to reproduce. Several more precise questions may be asked in this respect.

*Dimension of the spots*.—The diameter of the dark roundish spots was estimated to about 1 mm. Such a diameter is smaller than that of the vitellus of mature eggs of the genera *Alytes*, *Pelobates*, and *Pelodytes* laid by adults during reproduction, which are respectively of 2.2–7.0 mm, 1.5–2.5 mm and 1.4–2.1 mm (van den Elzen 1976; Arnold and Burton 1978; Alcover et al. 1984; Tarkhnishvili 1993). Thus if the tadpoles belonged to one of these genera, the size of the spots would be smaller than that of normal mature eggs of these genera, and it is unlikely that such eggs would be able to develop normally into viable organisms. The dark spots could thus represent immature oocytes. The only case in which the round spots could correspond to mature eggs would be if the studied tadpoles belonged to the genus *Pelophylax*, as mature eggs of frogs of this

group are particularly small, with a vitellus diameter of 0.92–2.5 mm (Chambers 1908; Arnold and Burton 1978; Berger and Uzzell 1980; Tarkhnishvili 1993; Tsiora and Kyriakopoulou-Sklavounou 2002; Miaud and Muratet 2004). Reasons for exclusion of this genus are discussed above. Furthermore, ovulated eggs in the body of anurans are always densely packed together, which is not the case of the spots under study, which show much space between them in two of our three samples (Fig. 4, 5).

At metamorphosis, most anuran ovaries contain oogonia and primary diplotene oocytes of different sizes (Al-Mukthar and Webb 1971: *Xenopus laevis*; Lopez 1989: *Bombina orientalis* (Boulenger, 1890); Ogielska and Wagner 1990: *Pelophylax ridibundus*; Viertel and Richter 1999. Oocytes at metamorphosis reach 100–130 μm in *P. ridibundus* (Ogielska and Wagner 1990; Wagner and Ogielska 1990). After metamorphosis, oogenesis and vitellogenesis (Wallace 1978; Wassermann and Smith 1978) and folliculogenesis (Guraya 1978) continue in a species-typical manner until and throughout the adult reproductive life (Echeverría 1988: *Rhinella arenarum* [Heusel, 1867]; Wagner and Ogielska 1990: *P. ridibundus*; Van Gansen 1986). Vitellogenesis starts after the first hibernation, whereas the first mature oocytes can be observed just before the third hibernation (Wagner and Ogielska 1990: *P. ridibundus*).

In conclusion, these data suggest that the spots, if they correspond to female reproductive cells, could not be mature oocytes likely to be fertilised and to develop into tadpoles.

*Position of the spots*.—The dark spots are situated in a posterior and ventral position in the abdomens of the two tadpoles concerned. This is not the place where ovaries and eggs contained in them would be expected to occur, should ovaries be liable to develop in larval anurans. In adult anurans, ovaries are situated along the vertebral column, often hidden by the kidney, and at any rate in a more anterior and dorsal than posterior and ventral position, the posteroventral part of the abdomen being occupied, in adults like in tadpoles, by the intestine. One possibility would be that the spots represent eggs that were no longer included in the ovary, but had already been ovulated and were about to be laid. Egg-laying in frogs normally requires that the eggs go through the oviduct, where they are given a jelly-coat that will expand in water and protect the vitellus in the external environment. Some of the connections that appear between the spots in the group of spots that was possibly not included in a tadpole could be interpreted as traces of jelly strings, like those that exist in the genera *Alytes*, *Pelobates*, and *Pelodytes*. But this interpretation would not hold for the spots that appear inside the tadpoles, which also show such connections: as long as anuran eggs are included within the female's oviduct, their jelly coat remains very thin, and it only expands with the contact of water outside the female (Lee 1964; Duellman and Trueb 1985: 114).

The anatomical limitations mentioned above, i.e., the lack of space within the abdomen of tadpoles that makes it unlikely that an ovary could expand there while its oocytes grow, also hold for the oviduct. At metamorphosis, the ovi-

duct of frogs is a very thin translucent linear structure, the Müller's canal, which only expands and becomes convoluted at the time of sexual maturity, i.e., much later than metamorphosis (Bhaduri 1953; Bhaduri and Basu 1957; Dubois 1976; Wake 1979; Viertel and Richter 1999). It is difficult to imagine how such a struture, not to mention an ovary full of large eggs, could develop within the abdomen of a tadpole without leading to a change of shape of this abdomen.

*Stage of tadpoles.*—The two tadpoles showing dark spots in their abdomens are approximately at Gosner stages 36–38. All the 17 other tadpoles, without spots, are at identical developmental stages, but some of them have a larger size (Table 1), so that it cannot be argued that eggs are present in the oldest tadpoles. With a sex-ratio of 1 male / 1 female, the expected number of female tadpoles would be nine or ten, and the difference between this number and the observed number of two is highly significant ($\chi^2 = 9.1$; $0.001 < P < 0.005$). Thus, if eggs are indeed present in some tadpoles of this population, this would not appear as a regular phenomenon, as it does not seem to affect all female tadpoles. A related question is the absence of young developmental stages among tadpoles: if some tadpoles in this population were laying eggs at the time of fossilisation, and if these eggs had been fertilised, one would expect some of these eggs to have given birth to young, tiny tadpoles; but of course the latter may have escaped fossilisation easier than larger tadpoles.

*Other considerations.*—The egg hypothesis can be further complicated by other hypotheses, which appear even more difficult to test. It could be imagined that, if tadpoles can develop eggs within their abdomen while being devoid of oviducts for their evacuation, such tadpoles could lay their eggs in the external environment through ventral tearing or "explosion", as occurs in some invertebrates. In some lower vertebrates, the Petromyzontidae Bonaparte, 1832 and the Myxinidae Bonaparte, 1832 (Agnathes), the oviducts are absent: the mature eggs pass through abdominal pores, which connect the coelomic cavity to the cloaca and are expelled outside by a rupture of the cloaca (Gérard 1958).

As another possibility, the eggs could have been laid by a normal adult amphibian, and two tadpoles would have swallowed such eggs but kept them undigested despite their progression through their digestive ducts, which seems quite unlikely. A single case of adults swallowing eggs but not digesting them is known in anurans, in the now extinct genus *Rheobatrachus* (Tyler 1983), where "gastric brooding" used to occur, but nothing similar is known in tadpoles. Furthermore, tadpoles that feed on eggs by shallowing them as a whole usually exhibit morphological specialisations (see Suggestions section below).

The question may be raised whether the eggs, if eggs were involved, were indeed inside the tadpoles before fossilisation. Clutches of eggs of amphibians are special environments that often attract organisms as a shelter, providing a warmer place (according to the conditions, frog egg masses may be several degrees warmer than the surrounding water)

or a source of food: often aquatic insects, fishes or amphibians (adult newts or frogs) are found in clutches of frog eggs, especially in cold waters (AD, personal observations). The two tadpoles MNHN 1998.9173–9174 could have been fossilised within egg masses and the eggs become superimposed on their abdomens during fossilisation. All these hypotheses are quite unlikely, but they are mentioned here in order to point to the uncertainties that remain regarding the interpretation of these observations.

Finally, let us stress that if eggs were laid by paedogenetic tadpoles, this does not mean that these eggs could develop: for this, they would need to be fertilised, which would require either complete paedogenesis in male tadpoles of the same species, or fertilisation by adult males—except if, beside being paedogenetic, these extraordinary tadpoles were also parthenogenetic!

European anurans of the genera discussed above all have a reproductive mode involving an amplexus in which the male grasps the female either above the hindlimbs or behind the forelimbs. Such an amplexus is impossible in tadpoles which are either devoid of free forelimbs (as long as they are covered by the operculum) or with forelimbs too short for grasping the large, ovoid body of the female tadpole. In order to ensure that sperm produced by a male tadpole could fertilise the eggs produced by a female tadpole, a completely new behaviour would need to be established, as well as anatomical structures allowing ejection of the gametes into the aquatic habitat. For paedogenesis to be possible in anuran tadpoles, maturation of gonads is not sufficient, but morphological and behavioural changes would be necessary. Evidence is missing that such major changes ever occurred in anurans. This point is certainly the strongest argument against the possibility of efficient paedogenesis in anurans.

**Suggestions for future research**.—Although very common organisms over most of our planet, tadpoles have still attracted very little attention. The first book dealing exclusively with these very peculiar and exciting organisms was published only ten years ago (McDiarmid and Altig 1999). As acknowledged by this book, still a lot has to be learned about them. The present observations and the hypotheses they suggest lead us to propose some questions that could be tested in future studies of tadpoles of living amphibians.

(i) Could there be some variation in the development stage reached by the gonad and the oviduct in tadpoles, according to the taxon and the conditions? If in some cases the ovary and oocytes can show some growth, where are they situated in the abdominal cavity? Could this account for the observation reported here?

(ii) Would it be possible to act upon the gonad and oviduct development of anuran tadpoles by chemical factors, e.g., those known to facilitate artifical paedogenesis in urodelans? It is possible to stop the development of anuran tadpoles with aminothiazol, thiouracile or thiourea, or simply by a thyroidectomy, while allowing the growth of gonads (Delsol 1952: *Discoglossus pictus* Otth, 1837). Similarly, the

ethinyltestosterone (pregneninolone) given in aqueous solution to tadpoles exerts a strong inhibiting action on the thyroid, blocking the somatic larval development. However, the masculinising action of this hormone in strong doses allows the development, during larval life, of immature testes (without spermatozoids) and of associated structures (Wolffian ducts and seminal vesicles) (Gallien 1949: *Rana temporaria* Linnaeus, 1758).

In thyroidless larvae of *Lithobates sylvaticus* (LeConte, 1825) (formerly *Rana sylvatica*, Ranidae; see Frost et al. 2006) kept through the second season after the controls have metamorphosed, the ovaries became large and large oocytes developed, although maturation was not observed and oviducts did not develop (the animals were not sexually mature). The testes became mature and produced spermatozoa which escaped into the kidneys (Hoskins and Hoskins 1919).

In similar experiments using thyroidectomy in *Lithobates pipiens* (Schreber, 1782) (formerly *Rana pipiens*, Ranidae), Allen (1917, 1918) demonstrated that spermatogenesis proceeds normally and sexual maturity is attained in thyroidless male tadpoles that have retained larval characteristics long after the time of metamorphosis. The ovaries of the female thyroidless tadpoles increased in size with time until they had reached a volume well in excess of that of the young normal frogs killed shortly after metamorphosis. The oocytes showed a diameter double of that in the recently metamorphosed frogs. In these experiments, artificial paedogenesis was definitely produced: sexual maturity was reached in the male and it is probable that it would have been reached in the course of time in the female tadpoles.

Some giant tadpoles from natural populations in Latvia showed "well-developed eggs in ovaries" and "testes normally developed" but no hindlimbs and a strongly pigmented skin (Borkin et al. 1982).

Finally, in a recent paper, Rot-Nikcevic and Wassersug (2004) reported about thyroidless tadpoles of *Xenopus laevis* occurring spontaneously in laboratory breeding colonies. These giant tadpoles stopped their somatic development and never metamorphosed. They showed an advanced gonadal development. The testes contained sperm, and the ovaries mature eggs with yolk. However, these tadpoles developed excessive musculature (especially the axial muscle mass of the tail) and had a scoliosis which deformed the back in dorsal view. This is obviously not what we observed in the fossil tadpoles described here.

The absence of thyroid hormones, while the hypophysis is still secreting prolactin and growth hormones, stops the somatic development although the development of the germ-line continues. The dissociation of the development of germen and soma allows the gonads to have time to reach maturity as the tadpole's size increases (Rot-Nikcevic and Wassersug 2004). Therefore, the largest tadpoles are usually the oldest and show the most advanced gonadal maturity. As the fossil tadpoles containing the black spots presented here were not the largest ones, this suggests that they are probably not thyroidless tadpoles.

(iii) Can some tadpoles, in some conditions, swallow whole amphibian eggs, or vegetal seeds, and keep them undigested through the intestinal duct? Could such a phenomenon explain the observations reported about here?

Tadpoles feeding from unfertilised eggs from their own mother are known in six anuran families: Dendrobatidae Cope, 1865 (*Oophaga pumilio* [Schmidt, 1857]: Weygoldt 1980; Crump 1983), Hylidae Rafinesque, 1815 (Noble 1929; Dunn 1937; Taylor 1954; Crump 1983; Lannoo et al. 1987; Jungfer and Schiesari 1995), Leptodactylidae Werner, 1896 (*Leptodactylus pentadactylus* group species: Muedeking and Heyer 1976; Hero and Galatti 1990; Magnusson and Hero 1991; Silva et al. 2005), Mantellidae Laurent, 1946 (*Mantella laevigata* Methuen and Hewitt, 1913: Glaw et al. 2000; Heying 2001), Microhylidae Günther, 1858 (*Hoplophryne* Barbour and Loveridge, 1928: Noble 1929; Crump 1983) and Rhacophoridae Hoffman, 1932 ("*Philautus*" cf. *carinensis*: Crump 1983; *Kurixalus eiffingeri* [Boettger, 1895]: Ueda 1986; Kam et al. 1996). However, the tadpoles of *Oophaga pumilio* feed on the eggs by biting a hole through the jelly capsule and sucking up the contents (Weygoldt 1980), so that the eggs are not wholly engulfed and would not be found complete in the gut. Similarly, *Mantella laevigata* eats the conspecific eggs and so does not swallow them (Glaw et al. 2000; Heying 2001), and only fragments of eggs were found in the guts of *Hoplophryne* tadpoles (Noble 1929). Tadpoles of the *Leptodactylus pentadactylus* species group also feed on heterospecific eggs. *Leptodactylus labyrinthicus* (Spix, 1824) tadpoles are reported to swallow entire eggs of *Dendropsophus minutus* (Peters, 1872), *Physalaemus* cf. *fuscomaculatus* (Steindachner, 1864). and *P. cuvieri* Fitzinger, 1826 (Silva et al. 2005). In *Osteopilus brunneus* (Gosse, 1851), from 5 to 184 whole eggs can be found in the stomach of the tadpoles, and a few intact eggs even in the intestine (Lannoo et al. 1987). In *Isthmohyla zeteki* (Gaige, 1929), at least 31 conspecific eggs were found in a stomach of a 9 mm head-body length tadpole (Dunn 1937). In the stomach of an unidentified rhacophorid species ("*Philautus*" cf. *carinensis*), 13 frog eggs with a mean diameter of 1.43 mm were found (Wassersug et al. 1981). No data about the number and size of the engulfed eggs were provided for the case of *Anotheca spinosa* (Steindachner, 1864) tadpoles (Taylor 1954), but according to the drawings, whole eggs are clearly visible through the body wall and localised posteroventrally in the abdominal cavity, reminiscent of the structures observed in this study. Oophagous tadpoles usually exhibit morphological features which appear adaptive relative to this feeding habit, such as a few keratodonts arranged in a few rows, a powerful head musculature, a short gut and an enlarged stomach. All the cases of cannibalism just listed concern tadpoles which develop in tree holes or in bromeliads, small habitats almost completely devoid of nutritive matter. This is a different situation from that of the Turkish tadpoles here described, which apparently lived in large lakes or swamps where organic material was presumably abundant.

However, a few cases are known of opportunistic cannibalism in tadpoles in temporary puddles. This was reported in *Isthmohyla pseudopuma* (Günther, 1901) by Crump (1983). Tadpoles preying on clutches of other frog species have also been reported (Heusser 1970, 2000; Mutz 2005), but, although these papers did not provide details, it is very likely that the eggs are rasped and therefore destroyed before being swallowed.

# Conclusion

Two of the 19 fossil tadpoles from the Miocene of Turkey here reported show unusual groups of regular roundish black spots in the abdominal region. Several possible hypotheses regarding the origin of these spots are discussed. None is fully satisfactory. In the improbable case where these spots would correspond to eggs, this would be the first reported case of paedogenesis in anurans, a phenomenon which has been so far considered impossible mostly for anatomical reasons (i.e., absence of space in the abdominal cavity). In the future, it would be interesting to explore this possibility, both experimentally and by broad anatomical survey of many tadpoles, both of living species and in fossil material.

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# References

Alberch, P., Gould, S.J., Oster, G.F., and Wake, D.B. 1979. Size and shape in ontogeny and phylogeny. *Palaeobiology* 5: 296–317.

Alcover, J.A., Mayol, J., Jaume, D., Alomar, G., Pomar, G., and Jurado, J. 1984. Biologia i ecologia de les poblacions relictes de *Baleaphryne muletensis* a la muntanya mallorquina. *In*: H. Hemmer and J.A. Alcover (eds.), *Història Biològica del Ferreret*, 129–151. Editorial Moll, Ciutat de Mallorca.

Al-Mukhtar, K.A.K. and Webb, A.C. 1971. An ultrastructural study of primordial germ cells, oogonia and early oocytes in *Xenopus laevis*. *Journal of Embryology and Experimental Morphology* 26: 195–217.

Allen, B.M. 1917. The effects of thyroid removal upon the development of the gonads in the larvae of *Rana pipiens*. *Science* 46: 216–218. doi:10.1126/science.46.1183.216 PMid:17782269

Allen, B.M. 1918. The results of thyroid removal in the larvae of *Rana pipiens*. *Journal of Experimental Zoology* 24: 499–519. doi:10.1002/jez.1400240303

Arnold, E.N. and Burton, J.A. 1978. *Tous les reptiles et amphibiens d'Europe en couleurs*. 271 pp. Elsevier Séquoia, Paris.

Baran, I. and Atatür, M.K. 1998. *Turkish Herpetofauna (Amphibians and Reptiles)*. 214 pp. Republic of Turkey Ministry of Environment, Ankara.

Basoglu, M. and Özeti, N. 1973. Türkiye amfibileri. *Ege Üniversitesi Fen Fakültesi Kitaplar Serisi* 50: 1–155.

Berger, L. and Uzzell, T.M. 1980. The eggs of European water frogs (*Rana esculenta*-complex) and their hybrids. *Folia Biologica* 28: 3–25.

Bhaduri, J.L. 1953. A study of the urogenital system of Salientia. *Proceedings of the Zoological Society of Bengal* 6: 1–111.

Bhaduri, J.L. and Basu, S.L. 1957. A study of the urogenital system of Salientia, Part I. Ranidae and Hyperoliidae of Africa. *Annales du Musée Royal du Congo Belge, Sciences Zoologiques* 55: 1–56.

Borkin, L.J., Berger, L., and Günther, R. 1982. Giant tadpoles of water frogs within *Rana esculenta* complex. *Zoologica Poloniae* 29: 103–127.

Boschwitz, D. 1957. Thyroidless tadpoles of *Pelobates syriacus* Boettger H. *Copeia* 1957: 310–311. doi:10.2307/1439175

Boulenger, G.A. 1891. A synopsis of the tadpoles of the European batrachians. *Proceedings of the Zoological Society of London* 1891: 593–627.

Bradford, D.F. 1984. Temperature modulation in a high-elevation amphibian, *Rana muscosa*. *Copeia* 1984: 966–976. doi:10.2307/1445341

Breuil, M. 1981. Une anomalie de la métamorphose : la persistance d'un intestin spiralé chez une Grenouille verte adulte (Amphibiens, Anoures). *Bulletin de la Société Linnéenne de Lyon* 50: 322–327.

Briggs, D.E.G. 2003. The role of decay and mineralization in the preservation of soft-bodied fossils. *Annual Reviews of Earth and Planetary Sciences* 31: 275–301.

Chambers, R. 1908. Einfluss der Eigrösse und der Temperatur auf das Wachstum und die Grösse des Frosches und dessen Zellen. *Archiv für Mikroskopische Anatomie* 72: 607–661. doi:10.1007/BF02982397

Clément, G. 1999. The actinistian (Sarcopterygii) *Piveteauia madagascariensis* Lehman from the Lower Triassic of northwestern Madagascar: a redescription on the basis of new material. *Journal of Vertebrate Paleontology* 19: 234–242.

Crump, M.L. 1983. Opportunistic cannibalism by amphibian larvae in temporary aquatic environments. *American Naturalist* 121: 281–287. doi:10.1086/284058

de Sá, R.O. 1988. Chondrocranium and ossification sequence of *Hyla lanciformis*. *Journal of Morphology* 195: 345–355. doi:10.1002/jmor.1051950308

Delsol, M. 1952. Action du thiouracile sur les larves de Batraciens. Néoténie expérimentale. Rôle de l'hypophyse dans ce phénomène. *Archives de Biologie* 63: 279–392.

Díaz-Paniagua, C. 1988. Temporal segregation in larval amphibian communities in temporary ponds at a locality in SW Spain. *Amphibia-Reptilia* 9: 15–26. doi:10.1163/156853888X00161

Dubois, A. 1976. Les Grenouilles du sous-genre *Paa* du Népal (famille Ranidae, genre *Rana*). *Cahiers népalais. Documents, Paris, CNRS* 6: i–vi + 1–275.

Dubois, A. 1979. Néoténie et pédogenèse. A propos d'une anomalie du développement chez *Bombina variegata* (Amphibiens, Anoures). *Bulletin du Muséum National d'Histoire Naturelle* (4), 1 (A): 537–546.

Dubois, A. 1987. Neoteny and associated terms. *Alytes* 4: 122–130.

Duellman, W.E. and Trueb, L. 1985. *Biology of Amphibians*. 691 pp. Mc Graw-Hill, New York (1986).

Dunn, E.R. 1937. The amphibian and reptilian fauna of bromeliads in Costa Rica and Panama. *Copeia* 1937: 163–167. doi:10.2307/1436136

Echeverría, D.D. 1988. Oogenesis en las hembras juveniles de *Bufo arenarum* (Anura, Bufonidae). *Revista del Museo Argentino de Ciencias Naturales "Bernardino Rivadavia" e Instituto Nacional de Investigación de las Ciencias Naturales, Zoología* 15: 57–75.

Elinson, R.P., Remo, B., and Brown, D.D. 1999. Novel structural elements identified during tail resorption in *Xenopus laevis* metamorphosis: lessons from tailed frogs. *Developmental Biology* 215: 243–252. doi:10.1006/dbio.1999.9481 PMid:10545234

Estes, R., Špinar, Z.V., and Nevo, E. 1978. Early Cretaceous pipid tadpoles from Israel (Amphibia: Anura). *Herpetologica* 34: 374–393.

Frost, D.R., Grant, T., Faivovich, J., Bazin, R.H., Haas, A., Haddad, C.F.B., de Sá, R.O., Channing, A., Wilkinson, M., Donnellan, S.C., Raxworthy, C.J., Campbell, J.A., Blotto, B.L., Moler, P., Drewes, R.C., Nussbaum, R.A., Lynch, J.D., Green, D.M., and Wheeler, W.C. 2006. The amphibian tree of life. *Bulletin of the American Museum of Natural History* 297: 1–370. doi:10.1206/0003-0090(2006)297[0001:TATOL]2.0.CO;2

Gall, J.C. 1991. Les fossiles au corps mou. *La Recherche* 22: 20–27.

Gallien, L. 1949. Dissociation de la chronologie du développement larvaire chez le têtard de *Rana temporaria* L., soumis à l'action de doses élevées d'éthinyl-testostérone. *Comptes Rendus hebdomadaires des Séances de l'Académie des Sciences* 229: 1367–1368.

Gaudin, A.J. 1973. The development of the skull in the Pacific tree frog, *Hyla regilla*. *Herpetologica* 29: 205–218.

Gaudin, A.J. 1978. The sequence of cranial ossification in the California toad, *Bufo boreas* (Amphibia, Anura, Bufonidae). *Journal of Herpetology* 12: 309–318. doi:10.2307/1563611

Gérard, P. 1958. Organes reproducteurs. *In*: P.P. Grassé (ed.), *Traité de Zoologie Anatomie, Systématique, Biologie XIII. Agnathes et Poissons, anatomie, éthologie, systématique (deuxième fascicule)*, 1565–1583. Masson et Cie, Paris.

Glaw, F., Schmidt, K., and Vences, M. 2000. Nachzucht, Juvenilfärbung und Oophagie von *Mantella laevigata* im Vergleich zu anderen Arten der Gattung (Amphibia: Ranidae). *Salamandra* 36: 1–24.

Godfrey, S.J. 1992. Fossilized eggs from Illinois. *Nature* 356: 21.

Godfrey, S.J. 1995. Fossilized eggs from the Pennsylvanian of Illinois. *Ichnos* 4: 71–75.

Gosner, K.L. 1960. A simplified table for staging embryos and larvae with notes on identification. *Herpetologica* 16: 183–190.

Gould, S.J. 1977. *Ontogeny and Phylogeny*. 517 pp. Belknap Press, Cambridge, MA.

Grande, L. 1984. Paleontology of the Green River formation, with a review of the fish fauna, 2nd ed. *The Geological Survey of Wyoming, Bulletin* 63: i–xviii + 1–333.

Grillitsch, B., Grillitsch, H., Haupl, M., and Tiedemann, F. 1983. *Lurche und Kriechtiere Niederösterreichs*. 176 pp. Facultas-Verlag, Wien.

Grossenbacher, K. 1997. *Alytes obstetricans* (Laurenti, 1768). *In*: J.-P. Gasc, A. Cabela, J. Crnobrnja-Isailovic, D. Dolmen, K. Groosenbacher, P. Haffner, J. Lescure, H. Martens, J.P. Martínez Rica, H. Maurin, M.E. Oliveira, T.S. Sofianidou, M. Veith, and A. Zuiderwijk (eds.), *Atlas of Amphibians and Reptiles in Europe*, 94–95. Societas Europaea Herpetologica and Muséum National d'Histoire Naturelle, Paris.

Guraya, S.S. 1978. Maturation of the follicular wall of nonmammalian vertebrates. *In*: R.E. Jones (ed.), *The Vertebrate Ovary. Comparative Biology and Evolution*, 261–330. Plenum, New York.

Hanken, J. and Hall, B.K. 1984. Variation and timing of the cranial ossification sequence of the Oriental fire-bellied toad, *Bombina orientalis* (Amphibia, Discoglossidae). *Journal of Morphology* 182: 245–255. doi:10.1002/jmor.1051820302

Hanken, J. and Hall, B.K. 1988. Skull development during anuran metamorphosis. I. Early development of the first three bones to form the exoccipital, the parasphenoid, and the frontoparietal. *Journal of Morphology* 195: 247–256. doi:10.1002/jmor.1051950303 PMid:3379643

Hero, J.-M. and Galatti, U. 1990. Characteristics distinguishing *Leptodactylus pentadactylus* and *L. knudseni* in the central Amazon rainforest. *Journal of Herpetology* 24: 226–228. doi:10.2307/1564240

Heusser, H. 1970. Laich-Fressen durch Kaulquappen als mögliche Ursache spezifischer Biotoppräferenzen und kurzer Laichzeiten bei europäischen Froschlurchen (Amphibia, Anura). *Oecologia* 4: 83–88. doi:10.1007/BF00390615

Heusser, H. 2000. Kaulquappen fressen Laich und Larven europäischer Anuren (Amphibia). *Zeitschrift für Feldherpetologie* 7: 177–202.

Heyer, W.R. 1973. Ecological interactions of frog larvae at a seasonal tropical location in Thailand. *Journal of Herpetology* 7: 337–361. doi:10.2307/1562868

Heying, H.E. 2001. Social and reproductive behaviour in the Madagascan poison frog, *Mantella laevigata*, with comparisons to the dendrobatids. *Animal Behaviour* 61: 567–577. doi:10.1006/anbe.2000.1642

Hirsch, K.F. 1994. The fossil record of vertebrate eggs. *In*: S.K. Donovan (ed.), *The Palaeobiology of Trace Fossils*, 269–294. John Wiley and Sons, Chichester.

Hoskins, E.R. and Hoskins, M.M. 1919. Observations on thyroidless *Rana sylvatica* larvae kept through the second season of normal metamorphosis. *Anatomical Record* 16: 152.

Huang, H. and Brown, D.D. 2000. Prolactin is not a juvenile hormone in *Xenopus laevis* metamorphosis. *Proceedings of the National Academy of Sciences of the USA* 97: 195–199. doi:10.1073/pnas.97.1.195

Huang, H., Marsh-Armstrong, N., and Brown, D.D. 1999. Metamorphosis is inhibited in transgenic *Xenopus laevis* tadpoles that overexpress type III deiodinase. *Proceedings of the National Academy of Sciences of the USA* 96: 962–967. doi:10.1073/pnas.96.3.962

Jungfer, K.-H. and Schiesari, L.C. 1995. Description of a central Amazonian and Guianan tree frog, genus *Osteocephalus* (Anura, Hylidae), with oophagous tadpoles. *Alytes* 13: 1–13.

Kam, Y.-C., Chuang, Z.-S., and Yen, C.-F. 1996. Reproduction, oviposition-site selection, and tadpole oophagy of an arboreal nester, *Chirixalus eiffingeri* (Rhacophoridae), from Taiwan. *Journal of Herpetology* 30: 52–59. doi:10.2307/1564706

Kasapligil, B. 1981. Past and present oaks of Turkey. Part I. *Phytologia* 49: 95–146.

Kemp, N.E. and Hoyt, J.A. 1969. Sequence of ossification in the skeleton of growing and metamorphosing tadpoles of *Rana pipiens*. *Journal of Morphology* 129: 415–444. doi:10.1002/jmor.1051290404 PMid:5353236

Kuzmin, S.L. 1997. *Pelodytes caucasicus* (Boulenger, 1896). *In*: J.-P. Gasc, A. Cabela, J. Crnobrnja-Isailovic, D. Dolmen, K. Groosenbacher, P. Haffner, J. Lescure, H. Martens, J.P. Martínez Rica, H. Maurin, M.E. Oliveira, T.S. Sofianidou, M. Veith, and A. Zuiderwijk (eds.), *Atlas of Amphibians and Reptiles in Europe*, 114–115. Societas Europaea Herpetologica and Muséum National d'Histoire Naturelle, Paris.

Lannoo, M.J., Townsend, D.S., and Wassersug, R.J. 1987. Larval life in the leaves: Arboreal tadpole types, with special attention to the morphology, ecology, and behavior of the oophagous *Osteopilus brunneus* (Hylidae) larva. *Fieldiana Zoology, N.S.* 38: i–iv + 1–31.

Lee, T.W. 1964. Maintenance of the unswollen state of the egg jelly in the *Rana pipiens* reproductive tract. *Journal of the Elisha Mitchell Science Society* 80: 1–8.

Llorente, G.A., Arano, B., Garcia-Serra, N., and Civantos, E. 1995. Extreme variability in the oral morphs of *Rana perezi* larvae: their dubious application in the diagnosis of the P-RP system. *Revista Española de Herpetología* 9: 85–91.

Lopez, K. 1989. Sex differentiation and early gonadal development in *Bombina orientalis* (Anura: Discoglossidae). *Journal of Morphology* 199: 299–311. doi:10.1002/jmor.1051990306

Maglia, A.M. and Púgener, L.A. 1998. Skeletal development and adult osteology of *Bombina orientalis* (Anura: Bombinatoridae). *Herpetologica* 54: 344–363.

Magnusson, W.E. and Hero, J.M. 1991. Predation and the evolution of complex oviposition behaviour in Amazon rainforest frogs. *Oecologia* 86: 310–318. doi:10.1007/BF00317595

Mamay, S.H. 1994. Fossil eggs of probable piscine origin preserved on Pennsylvanian *Sphenopteridium* foliage from the Kinney Quarry, central New Mexico. *Journal of Vertebrate Paleontology* 14: 320–326.

Mamay, S.H., Hook, R.W., and Notton, N., III. 1998. Amphibian eggs from the Lower Permian of north-central Texas. *Journal of Vertebrate Paleontology* 17: 80–84.

Martill, D.M. 1993. *Fossils of the Santana and Crato Formations, Brazil*. 158 pp. The Palaentological Association, London.

Maus, M. and Wuttke, M. 2002. Comparative anatomical and taphonomical examination of the larvae of *Pelobates decheni* Troschel 1861 and *Eopelobates anthracinus* Parker 1929 (Anura: Pelobatidae) found at the Upper Oligocene sites at Enspel (Westerwald/Germany) and Rott (Siebengebirge/Germany). *Courier Forschungsinstitut Senckenberg* 237: 129–138.

McDiarmid, R.W. and Altig, R. (eds.) 1999. *Tadpoles. The Biology of Anuran Larvae*. i–xiv + 1–444. University of Chicago Press, Chicago.

Miaud, C. and Muratet, J. 2004. *Identifier les œufs et les larves des amphibiens de France*. 200 pp. INRA Editions, Paris.

Muedeking, M.H. and Heyer, W.R. 1976. Descriptions of eggs and reproductive patterns of *Leptodactylus pentadactylus* (Amphibia: Leptodactylidae). *Herpetologica* 32: 137–139.

Mutz, T. 2005. Laichprädation durch die Kaulquappen des Mittelmeerlaubfrosches (*Hyla meridionalis*). *Zeitschrift für Feldherpetologie* 12: 260–265.

Nel, A. and Paicheler, J.-C. 1993. Les Libellulidae fossiles. Un inventaire critique (Odon., Anisoptera, Libelluloidea). *Entomologica Gallica* 4: 166–190.

Noble, G.K. 1929. The adaptive modifications of the arboreal tadpoles of *Hoplophryne* and the torrent tadpoles of *Staurois*. *Bulletin of the American Museum of Natural History* 58: 291–334.

Nöllert, A. and Nöllert, C. 1992. *Die Amphibien Europas*. 382 pp. Franckh-Kosmos, Stuttgart.

Ogielska, M. and Wagner, E. 1990. Oogenesis and development of the ovary in European green frog, *Rana ridibunda* (Pallas) I. Tadpole stages until metamorphosis. *Zoologische Jahrbücher Abteilung für Anatomie und Ontogenie der Tiere* 120: 211–221.

Olivier, E. 1893. Un crapaud phénomène. *Revue Scientifique du Bourbonnais et du Centre de la France* 6: 105.

Paicheler, J.-C. 1988. *Paléoécologie des milieux lacustres oligo-miocènes de la vallée du Gürçü-Dere (Anatolie-Turquie)*. 3 volumes, 465 pp. Unpublished Ph.D. thesis, Université de Reims, Reims.

Paicheler, J.-C., Broin, F. de, Gaudant, J., Mourer-Chauviré, C., Rage, J.-C., and Vergnaud-Grassini, C. 1978. Le bassin lacustre Miocène de Bes-Konak (Anatolie, Turquie): géologie et introduction à la paléontologie des vertébrés. *Géobios* 11: 43–65. doi:10.1016/S0016-6995(78)80018-7

Paicheler, J.-C., Nel, A., Gall, J.-C., and Delclòs, X. 2007. The aquatic genus *Notonecta* (Insecta: Heteroptera) as a palaeo-ecological indicator of rhythmite miming sequences in shallow freshwater deposits. *Journal of Taphonomy* 5: 105–120.

Pierce, B.A. and Smith, H.M. 1979. Neoteny or paedogenesis? *Journal of Herpetology* 13: 119–121. doi:10.2307/1563766

Rage, J.-C. and Roček, Z. 2003. Evolution of anuran assemblages in the Tertiary and Quaternary of Europe, in the context of paleoclimate and palaeogeography. *Amphibia-Reptilia* 24: 133–167. doi:10.1163/156853803322390408

Richardson, L.R. and Barwick, R.E. 1957. Faulty eruption of the fore-limb in *Hyla aurea*. *Transactions of the Royal Society of New Zealand* 84: 941–942.

Rot-Nikcevic, I. and Wassersug, R.J. 2004. Arrested development in *Xenopus laevis* tadpoles: how size constrains metamorphosis. *Journal of Experimental Biology* 207: 2133–2145. doi:10.1242/jeb.01002 PMid:15143146

Rückert-Ülkümen, N., Maus, M., Glaw, F., and Franzen, M. 2002. Kaulquappen von *Pelobates* sp. (Amphibia: Pelobatidae) aus dem Miozän von Beşkonak Köyü, Zentralanatolien, Türkei. *Mitteilungen der Bayerischen Staatssammlung für Paläontologie und historischen Geologie* 42: 75–82.

Sanchíz, B. 1998. Salientia. *In*: P. Wellnhofer (ed.), *Handbuch der Paläoherpetologie, Teil 4*. 287 pp. Verlag Dr. Friedrich Pfeil, München.

Savage, R.M. 1952. Ecological, physiological and anatomical observations on some species of anuran tadpoles. *Proceedings of the Zoological Society of London* 122: 467–514.

Silva, W.R., Giaretta, A.A., and Facure, K.G. 2005. On the natural history of the South American pepper frog, *Leptodactylus labyrinthicus* (Spix, 1824) (Anura: Leptodactylidae). *Journal of Natural History* 39: 555–566. doi: 10.1080/00222930410001671273

Špinar, Z.V. 1972. *Tertiary Frogs From Central Europe*. 286 pp. Junk, The Hague.

Tarkhnishvili, D.N. 1993. Anurans of Borjomi Canyon: clutch parameters and guild structure. *Alytes* 11: 140–154.

Taylor, E.H. 1954. Frog-egg-eating tadpoles of *Anotheca coronata* (Stejneger) (Salientia, Hylidae). *University of Kansas Science Bulletin* 36: 589–595.

Trueb, L. and Hanken, J. 1993. Skeletal development in *Xenopus laevis* (Anura: Pipidae). *Journal of Morphology* 214: 1–41. doi:10.1002/jmor.1052140102 PMid:1433306

Tsiora, A. and Kyriakopoulou-Sklavounou, P. 2002. Female reproductive cycle of the water frog *Rana epeirotica* in northwestern Greece. *Amphibia-Reptilia* 23: 269–280. doi:10.1163/15685380260449153

Tyler, M.J. (ed.) 1983. *The Gastric Brooding Frog*. 173 pp. Croom Helm, London.

Ueda, H. 1986. Reproduction of *Chirixalus eiffingeri* (Boettger). *Scientific Report of the Laboratory for Amphibian Biology, Hiroshima University* 8: 109–116.

van den Elzen, P. 1976. Remarques sur la biologie de *Pelodytes punctatus* Daudin 1802 (Amphibia, Pelobatidae) en Camargue. *Revue française d'Aquariologie* 3: 99–104.

Van Gansen, P. 1986. Ovogenèse des amphibiens. *In*: P.P. Grassé and M. Delsol (eds.), *Traité de Zoologie, Anatomie, Systématique, Biologie XIV, Fasc. 1B, Amphibiens*, 21–55. Masson et Cie, Paris.

Viertel, B. and Richter, S. 1999. Anatomy, viscera and endocrines. *In*: R.W. McDiarmid and R. Altig (eds.), *Tadpoles. The Biology of Anuran Larvae*, 92–148. The University of Chicago Press, Chicago.

Wagner, E. and Ogielska, M. 1990. Oogenesis and development of the ovary in European green frog, *Rana ridibunda* (Pallas). II. Juvenile stages until adults. *Zoologische Jahrbücher Abteilung für Anatomie und Ontogenie der Tiere* 120: 223–231.

Wake, M.H. 1979. The comparative anatomy of the urogenital system. *In*: H.M. Wake (ed.), *Hyman's Comparative Vertebrate Anatomy, 3rd ed.*, 555–614. University of Chicago Press, Chicago.

Wallace, R.A. 1978. Oocyte growth in nonmammalian vertebrates. *In*: R.E. Jones (ed.), *The Vertebrate Ovary. Comparative Biology and Evolution*, 469–502. Plenum, New York.

Wasserman, W.J. and Smith, L.D. 1978. Oocyte maturation in nonmammalian vertebrates. *In*: R.E. Jones (ed.), *The Vertebrate Ovary. Comparative Biology and Evolution*, 443–468. Plenum, New York.

Wassersug, R.J. 1974. Evolution of anuran life cycles. *Science* 185: 377–378. doi:10.1126/science.185.4148.377 PMid:4834372

Wassersug, R.J. 1975. The adaptive significance of the tadpole stage with comments on the maintenance of complex life cycles in anurans. *American Zoologist* 15: 405–417.

Wassersug, R.J., Frogner, K.J., and Inger, R.F. 1981. Adaptations for life in tree holes by rhacophorid tadpoles from Thailand. *Journal of Herpetology* 15: 41–52. doi:10.2307/1563645

Wassersug, R.J and Wake, D.B. 1995. Fossil tadpoles from the Miocene of Turkey. *Alytes* 12: 145–157.

Weygoldt, P. 1980. Complex brood care and reproductive behavior in captive poison-arrow frogs, *Dendrobates pumilio* O. Schmidt. *Behavioral Ecology and Sociobiology* 7: 329–332. doi:10.1007/BF00300674

Wiens, J.J. 1989. Ontogeny of the skeleton of *Spea bombifrons* (Anura, Pelobatidae). *Journal of Morphology* 202: 29–51. doi:10.1002/jmor.1052020104