

# Late Ordovician scolecodonts and chitinozoans from the Pin Valley in Spiti, Himachal Pradesh, northern India

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The end of the Ordovician witnessed major perturbations in the ecosystem, seriously affecting global marine biodiversity. Nevertheless, some marine organism groups and their crisis-bound palaeogeographic distribution are still understudied. Among the outliers are eunicid polychaetes, even though they flourished and diversified extensively during the Ordovician. A collection of seven genera of jaw-bearing polychaetes, including the new ramphoprionid genus *Spitiprion* Tonarová, Suttner, & Hints, with type new species of *Spitiprion khannai* Tonarová, Suttner, & Hints, is described here from Katian (Upper Ordovician) deposits of Spiti, northern India. The new species is preserved as isolated maxillae and a jaw cluster, and 3D models of the maxillary apparatus are reconstructed based on submicron-CT. Along with the scolecodonts, a low-diversity assemblage of chitinozoans was recovered, comprising five genera. The most common chitinozoan species are *Acanthochitina* cf. *cancellata* and *Spinachitina suecica*.

**Key words:** Polychaeta, Eunicida, Ramphoprionidae, Chitinozoa, organic-walled microfossils, Early Palaeozoic, Gondwana, submicron-CT.

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## Introduction

The Ordovician Period was a time of prominent biotic changes. The increase in biodiversity during the Ordovician has often been called the “Great Ordovician Biodiversification Event” (GOBE) (Webby et al. 2004; Servais et al. 2010; Ontiveros et al. 2023). However, nowadays, there are various opinions on the characteristics and timing of Ordovician radiation. This widely used term has been re-evaluated as various groups of organisms were characterised by different

diversification scenarios, and the process was not a short-lived event (Servais et al. 2023). Nevertheless, without a doubt, the end of the Ordovician was characterised by prominent changes in the atmosphere and oceans, including a period of glaciation (e.g., Brenchley et al. 2003) and a major perturbation in the carbon and sulphur cycling during the early Hirnantian (Schmidt et al. 2016) followed by a warming during the late Hirnantian (Bond and Grasby 2020). Just before the Hirnantian glaciation, another warming episode, the late Katian Boda Event, was documented through the

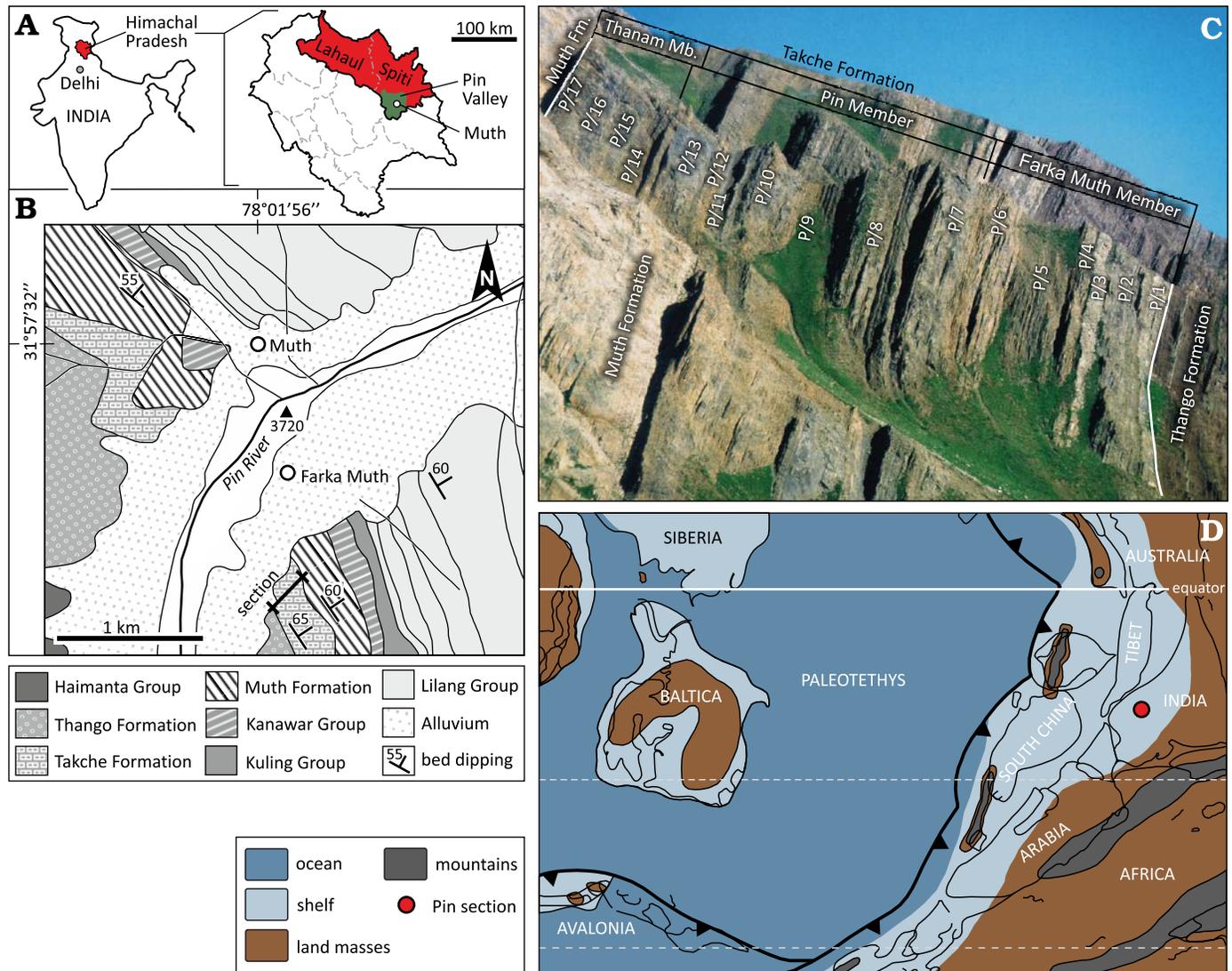


Fig. 1. A. Geographic location of the studied area within India. The state of Himachal Pradesh and an enlarged sketch of it with the Lahaul and Spiti district (red) and the Pin Valley (green). B. Geological map of the outcropping area within the Pin Valley in Spiti with the studied section south of Farka Muth indicated. The age of the lithostratigraphic units largely follows Bhargava and Bassi (1998): Haimanta Group (late Proterozoic to middle Cambrian), Sanugba Group including the Thango Formation (Ordovician) and the Takche Formation (Upper Ordovician to lower Silurian), Kanawar Group (Lower Devonian to lower Permian) with the Lower Devonian Muth Formation at its base, Kuling Group (Permian), Lilang Group (Lower Triassic to Upper Jurassic). C. Photograph of the studied section including the regional lithostratigraphic subdivision to units P/1–P/17. D. Palaeogeographic reconstruction with the assumed position of the Pin River section during the Late Ordovician (redrawn from Scotese 2000).

palaeogeographical shifts in shelly faunas and the nature of carbonate precipitation (Fortey and Cocks 2005; Suttner et al. 2007a; Lefebvre et al. 2014; Kröger et al. 2016; Myrow et al. 2019; Lu et al. 2021). However, most of our data on these environmental and biotic events comes from Laurentia and Baltica. More information is needed from other paleogeographic domains, including Gondwana.

The fossil record of soft-bodied invertebrates like polychaetes is rather uneven. Although body fossils have been reported from Palaeozoic strata, many lack diagnostic features of living taxa; hence, their proper taxonomic assignment is often impossible. The bulk of the fossil record of polychaetes is represented by their resistant jaws, the scolecodonts. The oldest finds of scolecodonts date back to the latest Cambrian

(Williams et al. 1999) and starting from the mid-Ordovician, they become widespread and diverse (Hints et al. 2017). The majority of preserved Early Palaeozoic jawed polychaetes belong to aciculate polychaetes, the Order Eunicida, a group of the subclass Errantia. They are characterised by the presence of reversible jaw apparatus, comprising ventral mandibles and dorsal maxillae in the ventral muscularised pharynx (Zanol et al. 2021). The Order Eunicida includes seven extant and 17 extinct families (Budaeva and Zanol 2021). Today, they are common in all types of marine environments (Fauchald 2013), inhabiting soft and hard substrates from intertidal to deep sea in all oceans; many burrow into hard corals and calcareous algae or live in their crevices (Hutchings 1986; Zanol et al. 2021).

The present paper aims to report and discuss a new Late Ordovician jawed polychaete fauna from the Takche Formation of the Pin River section near Farka Muth in the Pin Valley, Spiti, northern India. To date, only a few scolecodont maxillary apparatuses and isolated jaws are described from Gondwana, e.g., from the South African Soom Shale Lagerstätte (Whittle et al. 2008) or the Prague Basin (Tonarová et al. 2023). In addition to scolecodonts, we report the assemblage of chitinozoans, which were extracted from the same samples. The discussion of the chitinozoan assemblage contributes to the recently published palynological study from a coeval section near Gechang in the Parahio River Valley, Spiti, by Wang et al. (2021).

*Institutional abbreviations.*—CEITEC, Central European Institute of Technology, University of Technology, Brno, Czech Republic; CGS, Czech Geological Survey, Prague, Czech Republic.

*Other abbreviations.*—AL, attachment lamella; CT, computed tomography; M, maxilla.

*Nomenclatural acts.*—This published work and the nomenclatural acts it contains have been registered in Zoobank: urn:lsid:zoobank.org:pub:2CBBF838-A505-4F28-A745-13F78D379733

## Geological setting

Spiti, of which the Pin Valley is a part of, lies in the north Indian state of Himachal Pradesh and displays a well-exposed approx. 10 km thick sedimentary sequence ranging in age from the Neoproterozoic to Cretaceous (Bagati 1990; Bhargava and Bassi 1998; Fig. 1). The Ordovician–Silurian interval of the sequence is part of the Sanugba Group (approx. 1500 m thick) and divided into the Thango and Takche formations (compare Bhargava et al. 1991; Bhargava and Bassi 1998). A recently published summary of the lithostratigraphy of the ?Lower Ordovician–lower Silurian interval by Shabbar et al. (2023) shows that historically, the Thango and Takche units have also been called the Shian Formation and the Pin Formation, respectively (Srikantia 1974, 1977; Srikantia and Bhargava 2018 vs. Goel and Nair 1977, 1982; Suttner et al. 2001; Suttner 2003, 2007). To avoid confusion by parallel terminology for the Upper Ordovician–lower Silurian sequence in Spiti, the scheme of Shabbar and colleagues is followed here (Shabbar et al. 2023: fig. 3; the Takche Formation subdivided into the Farka Muth, Pin and Thanam members).

A Late Ordovician (Katian) to early Silurian (Llandovery) age of the middle and upper part of the Takche Formation (Pin and Thanam members) is constrained by conodonts in the Pin River and the Parahio River sections (Suttner 2003; Suttner et al. 2007b; Myrow et al. 2019). The siliciclastic lowermost unit (Farka Muth Member) has not provided conodonts to date. More promising are palyno-

morph studies on chitinozoans and acritarchs (e.g., Khanna and Sah 1983; Wang et al. 2021) or macrofossils, particularly brachiopods and trilobites (Reed 1912; Bhargava and Bassi 1998; Suttner 2003).

The rich and diverse fossil flora and fauna of Spiti have attracted scientists since the early–mid 19th century. The first stratigraphic discussions and monographs on early Palaeozoic fossils from Spiti were published by Salter and Blanford (1865), Stoliczka (1866), Griesbach (1891), Hayden (1904), and Reed (1912). Detailed stratigraphic documentation and taxonomic studies on calcimicrobes and calcareous algae were provided by Bhargava and Bassi (1986), Kato et al. (1987), Suttner and Hubmann (2005), Suttner et al. (2005), Hubmann and Suttner (2007), Pandey and Parcha (2018), Chaubey et al. (2019) and Vinn et al. (2023). Shabbar et al. (2022) focused on non-calcified warm-water macroalgae. Corals and stromatoporoids were reported by Bhargava and Bassi (1986, 1998), Suttner (2003), Suttner et al. (2007b) and Suttner and Kido (2011). Other fossil groups described include cephalopods (Suttner and Kröger 2006), ostracods (Schallreuter et al. 2008), cornulitids (Shabbar et al. 2023), bryozoans (Suttner and Ernst 2007) and conodonts (Suttner 2003; Suttner et al. 2007b; Myrow et al. 2019). Palaeoscoleoids, tentaculitids, gastropods, trilobites, brachiopods, and echinoderms were briefly characterised and figured by Reed (1912), Bhargava and Bassi (1998), Suttner (2003), and Suttner et al. (2007b).

Bhargava and Bassi (1986, 1998) studied five richly fossiliferous Ordovician–Silurian sections in detail. From the northwest to southeast, these are located within the Takche area, where the type section of the Takche Formation is exposed along the Spiti River bank (sensu Srikantia 1974, 1977 and Srikantia and Bhargava 2018), in the Parahio Valley near Gechang, in the Pin Valley near Farka Muth, at Leo and Manchap. Bed-by-bed lithological logs of the Takche Formation were provided only of the Pin River and Parahio River sections by Suttner (2003) and Myrow et al. (2019), respectively. The unit is generally characterised by a mixed siliciclastic-carbonate shallow-marine environment and varies in thickness between 145 and 280 m (Bhargava and Bassi 1986, 1998; Suttner 2003; Suttner et al. 2007b; Myrow et al. 2019; Chaubey et al. 2023).

Following the palaeogeographic reconstructions of Scotese (2000) and Torsvik and Cocks (2009, 2013), the area was a part of the Gondwana palaeocontinent and located between Arabia, Antarctica, and Australia (Fig. 1D). A palaeolatitudinal position at about ~25–30° S for northern India during the Late Ordovician–early Silurian, as suggested by Torsvik and Cocks (2009), is also supported by the occurrence of warm-water fossil groups like dasycladacean algae, stromatoporoids and corals in the Takche Formation.

*The Pin River Section.*—The studied section is located in the Pin Valley (Spiti) near the small village of Farka Muth at an altitude between ~3800–4100 m above sea level. The section can be reached by jeep or bus starting from Manali (Kullu) over Kaza, through Sangnam directly to Muth (GPS coor-

dinates: N 31°57'32", E 78°01'56") and Farka Muth (GPS coordinates: N 31°56'55", E 78°01'55"). Two high mountain passes (Rothang 3976 m and Kunzam La 4551 m) must be crossed before arriving at Losar on the border of the district (Suttner 2003). The sedimentary sequence of the Pin Valley was mapped in detail by, e.g., Fuchs (1982) and Bhargava and Bassi (1998). The section comprises early Palaeozoic sediments, including the uppermost portion of the Ordovician purple-coloured Thango Formation, followed by the grey Takche Formation, and succeeded by the white quartzites of the Lower Devonian Muth Formation (Fig. 1). A bed-by-bed lithological log of the 280 m thick Takche Formation at the Pin River section near Farka Muth was produced by Suttner (2003). Based on lithological characters, 17 distinct units (Figs. 1C, 2: P/1–P/17) were discriminated (Suttner 2003, 2007; Suttner et al. 2007b). Units P/1–P/6 consist of siliciclastic deposits and represent the Farka Muth Member (originally established by Suttner 2007). They are succeeded by a siliciclastic-influenced limestone portion (units P/7–P/13) which, according to Shabbar et al. (2022), represents the Pin Member. The upper portion of the Takche Formation (units P/14–P/17) is again siliciclastic and assigned to the Thanam Member (compare Shabbar et al. 2023: fig. 3). For the Pin Member, a Katian age is suggested by the occurrence of the Late Ordovician conodont *Amorphognathus ordovicicus* (Suttner 2003; Suttner et al. 2007b). Near the base of the Thanam Member, within the unit P/14, an early Silurian conodont fauna was identified (Suttner 2003; Suttner et al. 2007b). These biostratigraphic datings are also supported by a distinct carbon isotope excursion across the section (Suttner et al. 2007b).

## Material and methods

Initially, 41 rock samples were collected, processed by acid leaching (5% formic acid), wet sieved in four fractions (sieve mesh width: 63 µm, 125 µm, 250 µm, 500 µm), and dry-picked by TJS in 2000–2003 (Suttner 2003: 46–51, appendix). A further 16 samples (weight between 20–800 g) of the remaining collection stored in the repository of the Department of Palaeontology, University of Vienna, were dissolved to obtain additional scolecodonts. After dissolution in 6% acetic acid, the residues were gently rinsed through 80 and 50 µm sieves. The wet samples were handpicked for organic-walled microfossils from Petri dishes under a stereomicroscope, using a micropipette or an eyelash (wet technique sensu Kielan-Jaworowska 1966).

The insoluble residue contained scolecodonts, chitinozoans, tentaculitids, conodonts, ostracods, brachiopods, crinoids, gastropods, corals, silicified bryozoans, melanosclerites, spicules, and palaeoscolecid remains. Most invertebrate skeletons recovered after acid treatment were silicified or preserved as casts or moulds on clay-rich rock surfaces. In total, seven samples were productive for scolecodonts (P/7/31, P/8/27, P/8/69, P/11/80, P/13/117, P/13/231,

P/13/283), and six samples contained chitinozoans (P/7/31, P/8/69, P/8/78, P/12/9, P/13/117, P/13/231) (Fig. 2). The most productive sample for organic-walled microfossils was P/13/231. The preservation of microfossils varies from poor to very good. Some specimens could not be determined even to genus level due to flattening, deformation, or fragmentary preservation. The relatively small number of identifiable specimens did not allow a statistical analysis of the assemblage. The specimens were photographed using a Scanning Electron Microscope FEG-SEM Tescan Mira 3GMU at the Czech Geological Survey, Prague. X-ray micro- and submicron-CT were used for the study of internal structures and 3D reconstruction using the below-specified facilities of CEITEC (Brno). This technique is capable of non-destructive three-dimensional visualisation and analysis of the surfaces and internal structures of objects at a high spatial resolution. In CT, a series of X-ray projections is acquired over a circular arc, and cross-sections of the scanned samples are obtained from these projections by tomographic reconstruction. Several methods of mounting the specimens for the CT were tested. The best results were obtained using the following technique: the isolated specimens were prepared for measurement by placing them in a kapton tube and sealing the tube with parafilm. This preparation allowed the maxillae to be stable during CT measurements without any need for adhesives or other mounting measures. The maxillae were previously studied on SEM and therefore they were gold-coated. It turned out that the coating did not affect the study of internal structures, just there is a bright outer layer in the photos that was induced by the metal coating. In a few places, dark shadows appeared but did not influence the quality of measuring. Our study shows that the gold coating is not an obstacle for CT; it helped to make the specimens more resistant, which is useful when preparing the specimens for measurements. Preparation of the partly exposed scolecodont maxillary apparatus on a tiny piece of rock consisted of glueing it onto a rigid but low-density plastic rod. The cyanoacrylate glue fixed and stabilised the sample during measurement but did not interfere with parts of the sample containing the fossil material of interest.

The isolated maxillae were analysed by a Rigaku Nano3DX submicron-CT scanner (Rigaku Corporation, Japan). X-rays were produced using a MicroMax-007 HF generator operating at 50kV/24mA, with a molybdenum target. Projection images were acquired using a CCD (charge-coupled device) detector equipped with a scintillating crystal and magnifying microscope optics. Optical magnification was used because the quasi-parallel geometry of the nano3DX has very little geometric magnification. The rock piece with the jaw apparatus was studied by a Waygate Phoenix v|tome|x L240 micro-CT scanner (Waygate Technologies, Germany). A nanofocus transmission source at 60kV/120µA was used for X-ray production. Images were enlarged through geometric magnification and captured using a flat panel detector. Scan parameters for the measurements are summarised in

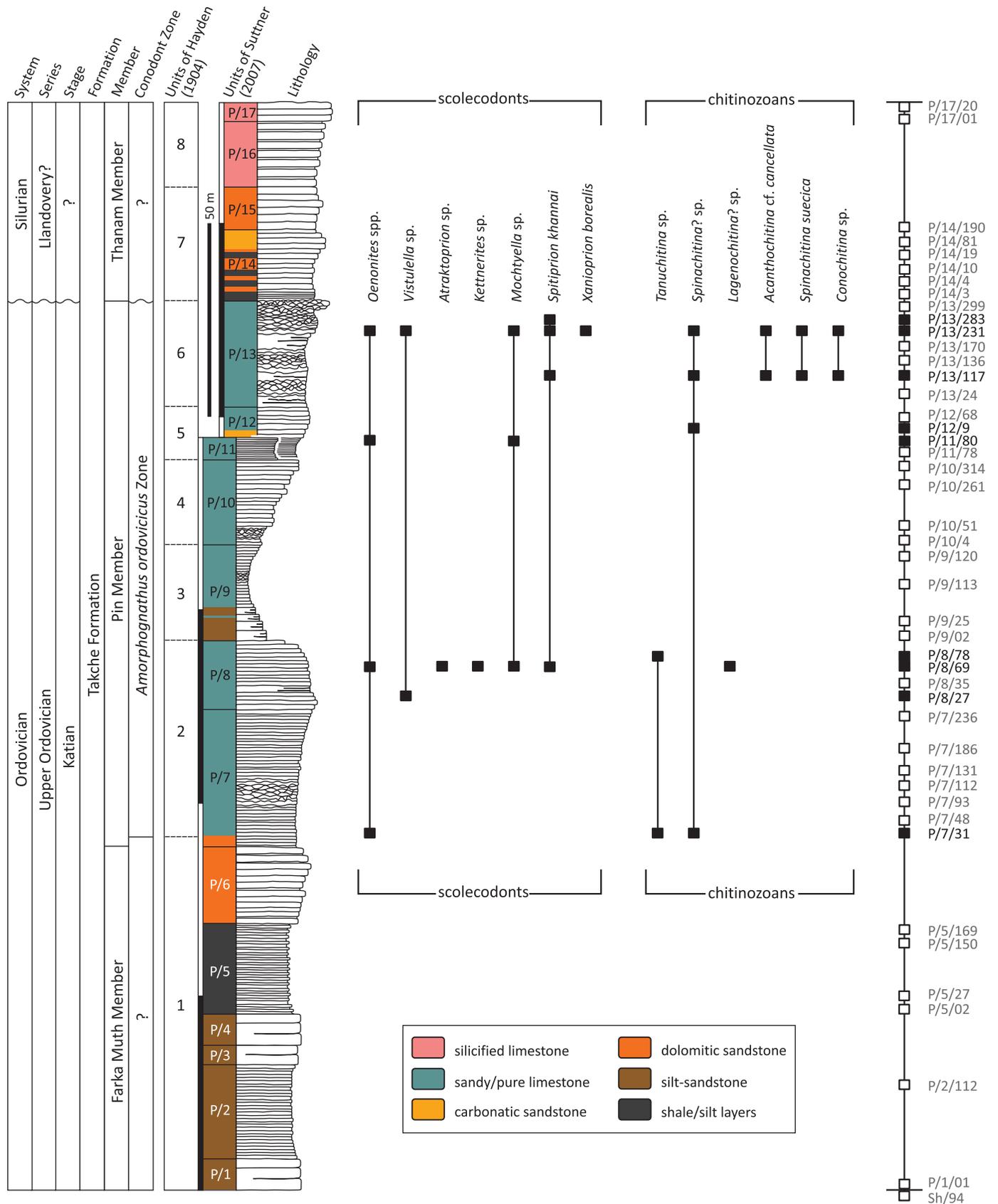


Fig. 2. Simplified lithological log of the Takche Formation at the Pin River section with the position of samples and the distribution of scolecodont and chitinozoan taxa. Sampling log on the right: solid squares denote productive samples for microfossils, whereas white squares indicate samples barren for scolecodonts and chitinozoans.

Table 1. CT scan parameters.

Sample	P13, isolated maxillae PT169_1 and PT169_2	Rock piece PT159_1
Scanner	nano3DX	v tome x L240
Tube voltage [kV]	50	60
Tube current [ $\mu$ A]	24000	120
Target material	Mo	W
Filter	0.1 mm Al	none
Linear voxel size [ $\mu$ m]	0.524	1.800
Scan range	180°	360°
Projection no.	800	2000 (3×average + 1 skip)
Exposure [s]	6	0.5

Table 1. The acquired data were reconstructed using the respective Rigaku and Waygate reconstruction software to obtain three-dimensional CT volumes. These were then processed using the software VGStudio MAX 2023.1 (Volume Graphics, Germany). Results in videos SOM 1–6 (Supplementary Online Material available at [http://app.pan.pl/SOM/app69-Tonarova\\_et\\_al\\_SOM.pdf](http://app.pan.pl/SOM/app69-Tonarova_et_al_SOM.pdf))

The sample code goes back to Suttner (2003) and is as follows: e.g., Pin River section/Unit 8/bed 69 = P/8/69. All figured specimens are housed in the collection of the Czech Geological Survey, collection numbers CGS PT156–170.

## Historical background

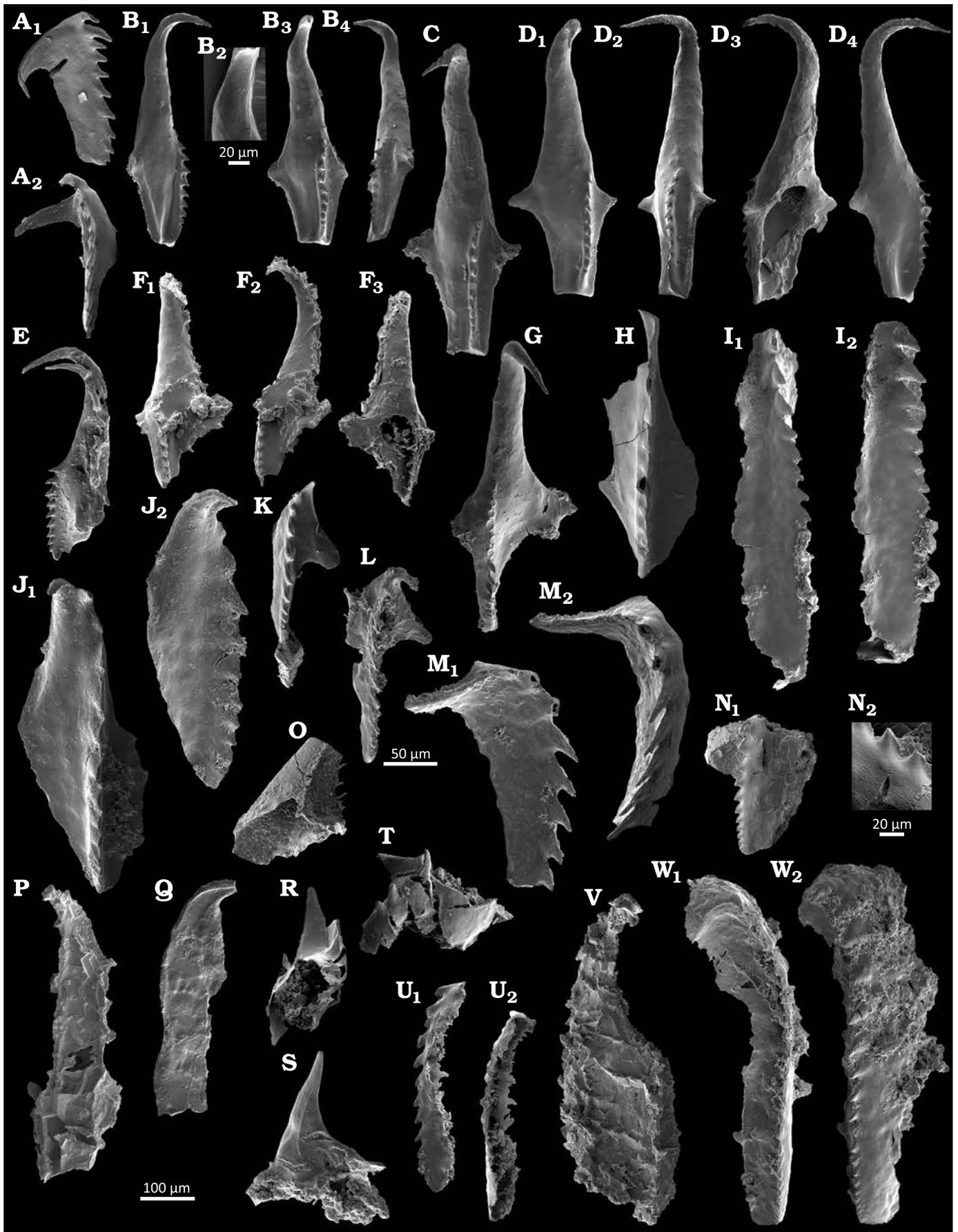
Most studies on Late Ordovician scolecodonts focused on Baltoscandia (Eriksson et al. 2013). Systematic studies were started already by Kielan-Jaworowska (1962, 1966) and, more recently, Hints (2000), Hints et al. (2010, 2023), and Hints and Tonarová (2023) provided analyses from the Baltica palaeocontinent. They concluded that the jawed polychaetes were diverse at that time, with a prominent component of mochtlyellids from the Placognatha group (*Pistoprion*, *Mochtlyella*, *Vistulella*) and polychaetaspids from the Labidognatha group (details on the concept of apparatus types can be found in Paxton 2009). The second best-covered area is Laurentia.

Late Ordovician scolecodonts from Laurentia were first studied e.g., by Stauffer (1933) and Eller (1942, 1945 and other papers of the author) or, more recently, by Bergman (1991), Eriksson and Bergman (2003), Eriksson et al. (2005), Schröer et al. (2016), or Hints et al. (2016). Overall, the faunas are dominated by labidognath and prionognath taxa, such as polychaetaspids, paulinitids, and atraktoprionids, with placognath taxa being less common than in Baltica. However, starting from the latest Ordovician, Laurentian and Baltic polychaete faunas have become increasingly similar (Eriksson et al. 2013).

There is only a handful of published records on Late Ordovician scolecodonts from Gondwana. These studies were summarised by Hints et al. (2015) and Tonarová et al. (2023). Hints et al. (2015) described a Late Ordovician assemblage from the Qusaiba-1 drill core, Saudi Arabia, and found out that most of the species recovered belong to the families Ramphoprionidae, Paulinitidae, Polychaetaspidae, and Atraktoprionidae, and only a very low proportion of taxa are with placognath type jaw apparatuses. Katian/Hirnantian scolecodonts from the Prague Basin (Levín locality) were studied by Tonarová et al. (2023). Similar to the Qusaiba-1 drill core, the taxa with labidognath and prionognath type maxillary apparatuses predominated in the Prague Basin samples, whereas placognath and ctenognath taxa were relatively rare. Polychaetaspids dominated the Levín assemblage, followed by other families such as ramphoprionids, paulinitids, and atraktoprionids. A similar general pattern is typical for the Laurentian samples but contrasts with the Baltic polychaete faunas.

Concerning India, the only previous report on scolecodonts was published by Khanna (1984), who worked on palynomorphs from lower Palaeozoic deposits of the “Pin Dolomite” (= Takche Formation) at the Parahio River section in Spiti. Khanna (1984) determined 15 single-element genera, but due to poor preservation of the material, proper identification was impossible. However, the presence of paulinitids and possibly mochtlyellids can be confirmed based on the specimens illustrated by Khanna (1984). Unfortunately, the work of Ashok Kumar Khanna remained unfinished due to a fatal accident during the fieldwork in 1984.

Fig. 3. Photomicrographs of eunicid polychaete scolecodonts from the Katian, Upper Ordovician of the Pin River section (Pin River Valley, Spiti, Himalaya, India). A–G. *Spitiprion khannai* gen. et sp. nov. A. CGS PT159\_2 (sample P/8/69), left MII in lateral (A<sub>1</sub>) and dorsal (A<sub>2</sub>) views. B. CGS PT159\_3 (sample P/8/69), left MI in lateral (B<sub>1</sub>, B<sub>4</sub>) and dorsal (B<sub>3</sub>) views, a close-up of the anterior part (B<sub>2</sub>). C. CGS PT159\_4, holotype (sample P/8/69), left MI in dorsal view. D. CGS PT169\_1 (sample P/13/231), left MI in dorsal (D<sub>1</sub>), dorsolateral (D<sub>2</sub>), and lateral (D<sub>4</sub>) views, ventral view (D<sub>3</sub>), note the polygonal structure in the myocoele opening. E. CGS PT170\_1 (sample P/13/283), right MI and lateral tooth in lateral view. F. CGS PT169\_2 (sample P/13/231), right MI in dorsal (F<sub>1</sub>), lateral (F<sub>2</sub>), and ventral (F<sub>3</sub>) views. G. CGS PT159\_5 (sample P/8/69), right MI in dorsal view. H. *Oeononites?* sp., CGS PT159\_6 (sample P/8/69), broken dorsal part of left MI in dorsal view. I. *Mochtlyella?* sp., CGS PT169\_3 (sample P13/231), right MI in dorsolateral view (I<sub>1</sub>), and in lateral view (I<sub>2</sub>). J–L. *Oeononites* sp. J. CGS PT169\_4 (sample P/13/231), left MI in dorsal (J<sub>1</sub>) and lateral (J<sub>2</sub>) views. K. CGS PT159\_7 (sample P/8/69), right MI in dorsal view. L. CGS PT169\_5 (sample P/13/231), right MI in dorsal view. M. *Xanioprion borealis* Kielan-Jaworowska, 1962, CGS PT169\_6 (sample P13/231), left MII in lateral (M<sub>1</sub>) and dorsal (M<sub>2</sub>) views. N, O, R–U. Scolecodonta indet. N. CGS PT159\_8 (sample P/8/69), fragment of an undetermined maxilla in lateral view (N<sub>1</sub>) and detail of the dentary (N<sub>2</sub>). O. CGS PT159\_9 (sample P/8/69), fragment of the lateral tooth. R. CGS PT159\_12 (sample P/8/69), single lateral tooth. S. CGS PT159\_13 (sample P/8/69), single lateral tooth. T. CGS PT159\_14 (sample P/8/69), single lateral tooth. U. CGS PT169\_7 (sample P/13/231), placognath maxilla in lateral (U<sub>1</sub>) and ventral (U<sub>2</sub>) views. P, Q. *Kettnerites* sp. P. CGS PT159\_10 (sample P/8/69), right MI in dorsal view. Q. CGS PT159\_11 (sample P/8/69), left MI in dorsal view. V. *Atraktoprion* sp., CGS PT159\_15 (sample P/8/69), left MI in dorsal view. W. *Vistulella* sp., CGS PT169\_7 (sample P/13/231), left MI in dorsal (W<sub>1</sub>) and dorsolateral (W<sub>2</sub>) views. The scale bar refers to all specimens except for B<sub>2</sub>, L, and N<sub>2</sub>, where a separate bar is indicated.



The Late Ordovician chitinozoans from Gondwana are far better studied than scolecodonts. Here, we mention only some examples of the previous studies that are more relevant to our new material. Firstly, several papers describe chitinozoan faunas from the Middle East, e.g., by Al-Hajri (1995) or Al-Shawareb et al. (2017) from the Katian of Saudi Arabia, Paris et al. (2015) from the Katian and Hirnantian of Saudi Arabia, Ghavidel-Syooki (2001, 2023), Ghavidel-Syooki and Winchester-Seeto (2002), Ghavidel-Syooki and Piri-Kangarshahi (2021) from Iran, among others. The South American chitinozoans were studied by Grahn (1992) in Brazil, de la Puente et al. (2020) and de la Puente and Astini (2023) in Argentina. Other reports come from Northern Africa, from Morocco by Elaouad-Debbaj (1984) and Bourahrouh et al. (2004), and from Libya by Paris (1988) and Thusu et al. (2013). South China is relatively well-studied (e.g. Liang et al. 2023), and the first reports from the Prague Basin were presented by Tonarová et al. (2023).

Within the Himalayas, Sinha et al. (2011) and Sinha and Verniers (2016) discovered Middle to Late Ordovician chitinozoans from the Shiala Formation of the Garhwal-Kumaon subbasin, Uttarakhand, India. Khanna and Sah (1983), Khanna (1984) and Khanna et al. (1985), however, were the first authors to mention the presence of chitinozoans and other palynomorphs in the Spiti area. Chitinozoans from the Parahio River section, representing the time interval before and during the Boda event, were studied by Wang et al. (2021). That section, measured bed-by-bed by Myrow et al. (2019), is located approximately 12 kilometres north of the Pin River section near Farka Muth, as reported in our study. Wang et al. (2021) found 13 species of chitinozoans belonging to seven genera, indicating a late Katian age.

## Systematic palaeontology

Scolecodont descriptive terminology is based on that of Kielan-Jaworowska (1966), Jansonius and Craig (1971) and Bergman (1989).

Class Polychaeta Grube, 1850

Order Eunicida Dales, 1963

Family Ramphoprionidae Kielan-Jaworowska, 1966

Genus *Spitiprion* Tonarová, Suttner, & Hints nov.

Zoobank LSID: urn:lsid:zoobank.org:act:560719CA-4C5D-47EB-B077-077546969C97

*Etymology*: After Pin River section Spiti in the Trans-Himalayan region of northern India.

*Type species*: *Spitiprion khannai* Tonarová, Suttner, & Hints sp. nov., see below. By monotypy.

*Diagnosis*.—As for the type species.

*Remarks*.—The closest known ramphoprionid to *Spitiprion* is *Megaraphoprion* Eriksson, 2001. The biggest difference is fully denticulated dentary of the first maxillae of the

latter genus and the ramus of right first maxilla (MI) is more slender in *Spitiprion*. *Spitiprion* has also much more prominent and slender hooks on the first maxillae than the type species *M. magnus*. The second maxillae of *S. khannai* Tonarová, Suttner, & Hints gen. et sp. nov. also differ considerably from the type species of *Megaraphoprion*. The second maxillae (MII) of *M. magnus* are sturdy, with wide ramus and blunt teeth whereas the MIIs of *S. khannai* Tonarová, Suttner, & Hints gen. et sp. nov. are rather slender, with pointed denticles and pointed ramus, reminding the second maxillae of *Ramphoprion* Kielan-Jaworowska, 1962.

*Stratigraphic and geographic range*.—Katian, Upper Ordovician of Pin Member (*Amorphognathus ordovicicus* Conodont Biozone), Takche Formation, Pin Valley, Spiti, India.

*Spitiprion khannai* Tonarová, Suttner, & Hints sp. nov.  
Figs. 3A–G, 4; SOM 1–6.

Zoobank LSID: urn:lsid:zoobank.org:act:7D98ABE7-300E-4D50-9028-9608403D9ABA

*Etymology*: In honour of the Indian palaeontologist Ashok K. Khanna (1951–1984), who first described scolecodonts from India.

*Holotype*: CGS PT\_159\_4, left first maxilla, from sample P/8/69.

*Type locality*: Pin River section, Spiti, India.

*Type horizon*: *Amorphognathus ordovicicus* Conodont Biozone, Katian, Upper Ordovician.

*Material*.—Five isolated left MI (CGS PT\_159\_3, 159\_4, 169\_1, 169\_16, 170\_2), 5 isolated right MI (CGS PT\_159\_5, 168\_8, 169\_2, 159\_17, 170\_1), 1 MII (CGS PT\_159\_2), 1 partly preserved apparatus (including left and right MI, MIV) (CGS PT\_159\_1). All from the type locality and horizon.

*Diagnosis*.—MI elongated, widest in the posterior third. The dentary spread along the posterior half of the jaw length, terminating with a prominent and bent anterior hook. The right MI with a conspicuous slender ramus of rectangular shape, with a prominent inner wing occupying approximately posterior third of jaw, the wing is ending with a pointed extension. The left MI with a prominent inner wing extending approx. posterior 40% of the jaw, ending by a prominent pointed extension. In the outer margin, slightly below the centre protrudes a prominent extension, symmetrically to the inner wing extension. The dentary of both first maxillae is occupied by 8 to 12 relatively small but sharp denticles. The un-denticulated hook starts at approx. mid-length of the jaw, and it has an almost whip-like ending. The myocoele opening occupies about one-third of the jaw length. The MIIs are expected to be near mirror images of each other, with a slender ramus placed in the anterior-most part, pointed denticles, with the most prominent first denticle. MIV (right confirmed) is slightly longer than wide, occupied with relatively large and sturdy denticles. The basal plate was not found, but most likely it is sub-rectangular, with small denticles and a tapering antero-dextral process.

*Description*.—*Right MI*: Length 480–540 µm, width 120–126 µm, width is approx. four times shorter than length

(width 0.2–0.3 of jaw length), dorsal view: the 9–12 pointed denticles are bent outwards, with the tip often broken off. The undenticulated hook extends for approx. half of the maxilla length (compared with other ramphoprionids that have denticulated also the anterior part of the dentary), the bend pointed tip is very slender and long. Typical for the species is a prominent extension of the maxilla on the centre of both outer and inner face.

Ventral view: the myocoele opening represents approx. 1/3 of the maxilla. In the ventral view, all denticles are visible through the myocoele opening (i.e., the length of the denticulated dentary approximately matches the length of the myocoele opening).

In the right MI (Fig. 3E) remained attached also an attachment lamella (AL). The ALs are attached to the bases of MI, and when the jaws are retracted, the AL extend anteriorly-posteriorly on either side of the maxillae to near MIII and MIV, therefore they may increase resistance to hyperextension of the primary maxillae that may result from contact of the open jaws with struggling prey or surrounding substrate—this potential function of the AL is consistent with the motile carnivorous lifestyle of *Lumbrinereis latreilli* Audouin & Milne Edwards, 1834 (Clemo and Dorgan 2017). However, it is supposed that ramphoprionids do not have descendants in the recent oceans (Kielan-Jaworowska 1966), therefore the lifestyle and diet remain speculative.

*Left MI*: Length 430–580  $\mu\text{m}$ , width 126–138  $\mu\text{m}$ , width is approx. 4 times shorter than length (width 0.2–0.3 of jaw length), dorsal view: the shape of the maxilla is rectangular in the posterior area and tapers anteriorly to the undenticulated anterior part that extends for half of the maxilla length and ends with a prominent hook-like slender fang. The dentary consists of ten pointed denticles that are situated only in the posterior half of the dentary, the denticles are bent outwards, and decrease in size toward the posterior end, with the smallest first two and last two denticles. Posteriormost denticles are tightly packed and continue into the undenticulated short ridge. The posterior margin is rather short, it extends 14% of the jaw length. Among other ramphoprionids the posterior margin represents 20 to 30% of the jaw length in *Protarabellites* species, 15 to 20% of the jaw length in *Ramphoprion* species, and 19 to 25% in *Megaramphoprion magnus* Eriksson, 2001. In ventral view, the myocoele opening extends a little more than 1/3 of the maxilla (approx. 35–40% of the jaw length), which is more closed than in other ramphoprionid species—where it is usually representing from 0.4 (*Ramphoprion bialatus* Hints, 1998) to 0.8 (*Protarabellites triangularis*) of the maxilla length.

*Left MII*: Length 300  $\mu\text{m}$ , width 180  $\mu\text{m}$  (0.6 of the jaw length). The dentary is prominent, strongly elevated, gently curved, occupying the entire jaw length, and equipped with ten denticles of similar size as in the first maxillae. The first two biggest teeth are followed by two smaller denticles and then a row of teeth of similar size. The ramus is prominent

but very narrow, triangular in shape. In the ventral view, the myocoele is almost gaping.

*Right MIV*: Jaw is about as high as long, it is quite flat, with an almost square outline. The left side is occupied by seven to eight sturdy denticles of more or less similar size, except for the first bigger denticle. The preservation (flattening) of MIV may have influenced the appearance of the maxilla and may be modified when more material is at hand.

Other jaws presently unknown.

*Remarks*.—The ramphoprionids, known solely by their jaws (Eriksson 2002), emerged in the Middle Ordovician (Kielan-Jaworowska 1962, 1966; Szaniawski 1996) and their youngest known record is late Ludfordian (Ludlow, Silurian) in age (Eriksson 2001). Up to now, there were four genera included in the family, the type genus *Ramphoprion* Kielan-Jaworowska, 1962, *Protarabellites* Stauffer, 1933, *Pararamphoprion* Männil & Zaslavskaya, 1985, and *Megaramphoprion* Eriksson, 2001. Eriksson (2002) suggested that the Silurian ramphoprionids were inhabiting only regions close to the equator, but Tonarová et al. (2012) confirmed their presence in the Ludfordian of the Prague Basin, which was located between 25 and 30 degrees south during the Ludfordian (Tasáryová et al. 2014).

The family Ramphoprionidae was considered basal and fairly primitive among labidognath families (Eriksson 2001). However, their jaw apparatuses exhibit characters considered both primitive, such as denticulation in first maxillae and advanced, such as enclosed to sometimes near strongly enclosed myocoeles. Therefore, ramphoprionids were presumably more advanced than the polychaetaspids, a group with similar jaw apparatus architecture, although the latter apparently have a longer (documented) stratigraphic range (cf. Kielan-Jaworowska 1966; Szaniawski 1996).

However, the first maxillae of *Spitiprion khannai* Tonarová, Suttner, & Hints gen. et sp. nov. differ from other ramphoprionids by the reduced denticulation to only half of the maxillae. Eriksson (2001: figs. 9.28, 9.31–44, and 45, and 46, respectively) reported from Gorstian and Ludfordian (Ludlow, Silurian) of Gotland a decrease in denticulation of the species *Ramphoprion gotlandensis* in the anterior part of the first maxillae dentaries, but that was reflected by a paucidate dentary, i.e., usually blunt denticles with longer gaps in between. The loss of denticulation in the first maxillae is observed in several extant polychaete groups, e.g., in the families Eunicidae or Onuphidae (Beesley et al. 2000; Paxton 2009). However, the denticulation can be variable, e.g., in the labidognath family Paulinitidae Lange, 1947 (*Langeites lublinensis* Szaniawski and Wrona, 1973) and in the extant prionognath family Oeonidae (*Drilonereis* Claparède, 1870). Palaeozoic paulinitids were considered directly ancestral to modern labidognaths by Kielan-Jaworowska (1966).

*Stratigraphic and geographic range*.—Katian, Upper Ordovician, Pin Member (*Amorphognathus ordovicicus* Conodont Biozone), Takche Formation, Pin Valley, Spiti, India.

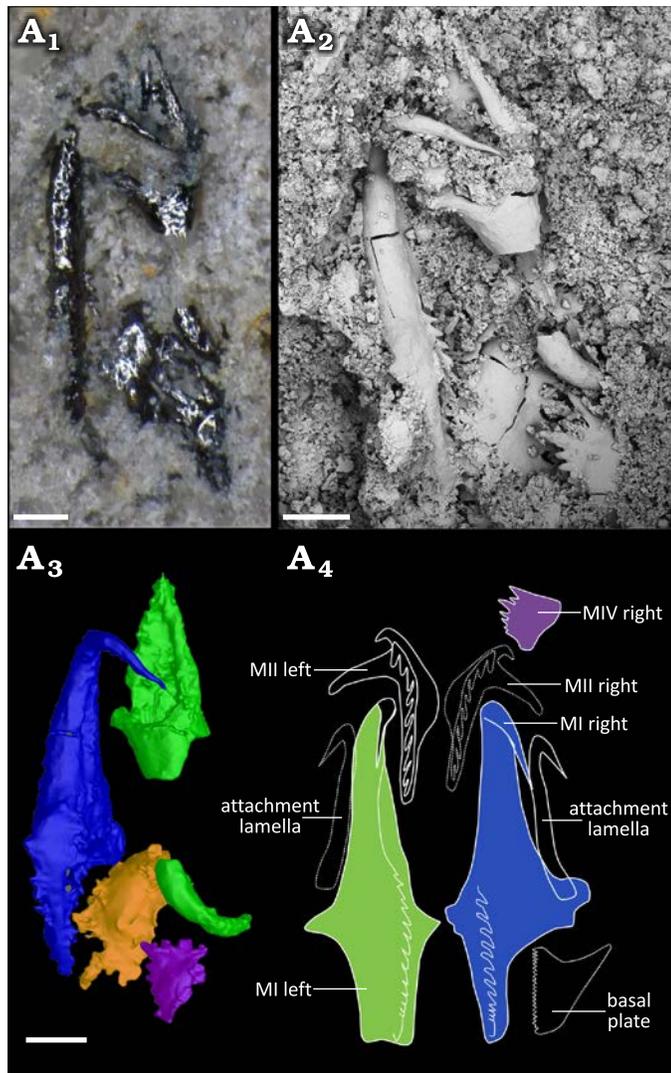


Fig. 4. Photomicrographs of the eunicid polychaete *Spitiopriion khannai* gen. et sp. nov. apparatus preserved on an acetic acid-etched rock surface, CGS PT159\_1 (sample P/8/69) from the Katian, Upper Ordovician of the Pin River section (Pin River Valley, Spiti, Himalaya, India). A<sub>1</sub>, optical microscope photograph; A<sub>2</sub>, SEM photomicrograph; A<sub>3</sub>, 3D reconstruction based on images from X-ray micro-computed tomography. The affinity of the maxilla (orange) is yet unknown (could be the second maxilla but diagnostic features are missing). A<sub>4</sub>, sketch of the maxillary apparatus of the new species of eunicid polychaete *Spitiopriion khannai*. The maxillae (white) with dotted lines are assumed according to closely related species but have not been found yet. MI, first maxilla; MII, second maxilla; MIV, fourth maxilla; AL, attachment lamella; BP, basal plate. Scale bars 100  $\mu$ m.

## Results

**Scolecodonts.**—The collection of scolecodonts from Katian deposits of the Takche Formation at the Pin River section near Farka Muth in Spiti includes approx. 60 maxillae (counting only first or second maxillae, which can be determined at least to the generic level). These were assigned to seven genera belonging to six families: Polychaetaspidae, Paulinitidae, Ramphoprionidae, Mochtyellidae, Xanioprionidae, and Atraktoprionidae (Figs. 2, 3). The maxillae are fragile and commonly fragmentarily preserved. Some of the jaws also reflect scars of mineral growth. All these conditions impede species-level identification. The two richest samples were P/8/69 and P/13/231, both contained representatives of five different polychaete genera (Fig. 2).

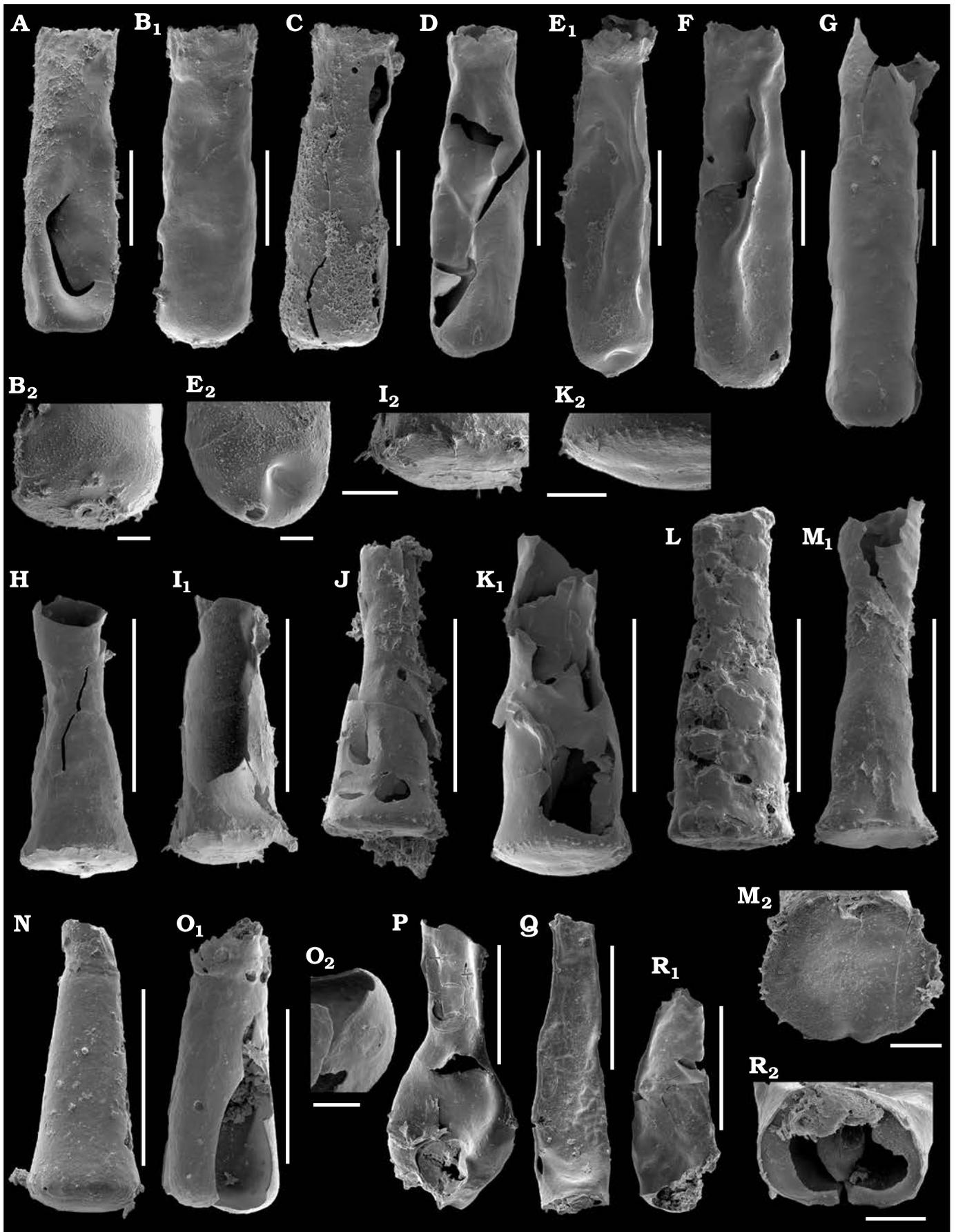
Ramphoprionids are represented by a new genus and species *Spitiopriion khannai* Tonarová, Suttner, & Hints. Representatives of the new species occur in units P/8 and P/13 of the Takche Formation at the Pin River section. Additionally, a maxillary apparatus consisting of four jaws (paired MI, MII and MIV) partly embedded in a piece of rock, was found in sample P/8/69 and studied using micro-CT (Fig. 4). The isolated maxillae of the new species from the sample P/13/231 were studied using a submicron-CT (Table 1, SOM 1–6; same specimens are on Fig. 3D and F, respectively; for a detailed description see Systematic palaeontology).

Jaws of polychaetaspids were found in three samples (P/7/31, P/8/69, P/11/80, P/13/231), and all of them are assigned to *Oeononites* Hinde, 1879, with at least two species present. Because of poor preservation, determination at the species level was impossible; thus, the specimens were assigned to *Oeononites* spp. (Fig. 3H, J–L).

Paulinitid maxillae were found only in one sample (P/8/69). The specimens have been determined as *Kettnerites* Žebera, 1935. However, the first maxillae of *Kettnerites* sp. (Fig. 3P, Q) are so distinctive that there is no doubt about their assignment.

Placognath taxa are relatively rare in the samples. Their saw-like maxillae tend to break, and therefore taphonomy may have biased the abundance quite significantly. The sample P/13/231 contains most of the placognath forms, but it is also the most productive sample overall. Among mochtyellids, two genera were recorded: *Vistulella* sp. (Fig. 3W) and *Mochtyella* sp. (Fig. 3I). Xanioprionids are also very rare in the Pin River section—only a single maxilla was recovered

Fig. 5. Photomicrographs of selected chitinozoans from the Katian, Upper Ordovician of the Pin River section (Pin River Valley, Spiti, Himalaya, India). A–G. *Acanthochitina* cf. *cancellata* Martin, 1983. A. CGS PT169\_9 (sample P/13/231). B. CGS PT169\_10 (sample P/13/231), general view (B<sub>1</sub>), close-up showing the base in detail (B<sub>2</sub>). C. CGS PT169\_11 (sample P/13/231). D. CGS PT169\_12 (sample P/13/231). E. CGS PT169\_13 (sample P/13/231), general view (E<sub>1</sub>), close up showing the base in detail (E<sub>2</sub>). F. CGS PT169\_14 (sample P/13/231). G. CGS PT168\_1 (sample P/13/117). H–M. *Spinachitina suecica* (Laufeld, 1967). H. CGS PT168\_2 (sample P/13/117). I. CGS PT168\_3 (sample P/13/117), general view (I<sub>1</sub>), close up showing the base in detail (I<sub>2</sub>). J. CGS PT168\_4 (sample P/13/117). K. CGS PT168\_5 (sample P/13/117), general view (K<sub>1</sub>), close up showing the base in detail (K<sub>2</sub>). L. CGS PT169\_15 (sample P/13/231). M. CGS PT168\_6 (sample P/13/117), general view (M<sub>1</sub>), close up showing the base in detail (M<sub>2</sub>). N. *Spinachitina?* sp., CGS PT156\_1 (sample P/7/31). O. *Conochitina* sp., CGS PT168\_7 (sample P/13/117), general view (O<sub>1</sub>), close-up of the base of the specimen with a mucron (O<sub>2</sub>). P. *Lagenochitina?* sp., CGS PT156\_16 (sample P/8/69). Q, R. *Tanuchitina* sp., Q. CGS PT156\_2 (sample P/7/31). R. CGS PT156\_3 (sample P/7/31), general view (R<sub>1</sub>), close-up showing the base in detail and the short carina (R<sub>2</sub>). Scale bars 100  $\mu$ m, except for enlargements B<sub>2</sub>, E<sub>2</sub>, I<sub>2</sub>, K<sub>2</sub>, M<sub>2</sub>, O<sub>2</sub>, R<sub>2</sub> 20  $\mu$ m.



and identified as *Xanioprion borealis* Kielan-Jaworowska, 1962 (Fig. 3M).

Additionally, a single maxilla from sample P/8/69 was assigned to *Atraktoprion* sp. (Fig. 3V), showing close similarity to *Atraktoprion* sp. A of Eriksson et al. (2005).

**Chitinozoans.**—The collection of chitinozoans from the Pin River section includes approx. 50 specimens, which are assigned to five genera (Fig. 2). Generally, the specimens are characterised by their black vesicles. The appendices are commonly broken off, and the ornamentation is difficult to recognise. Most abundant are specimens with a medium conical test of about 300–400 µm in length, characterised by distinct, web-like, regularly distributed spines (Fig. 5C) suggesting mesh-like ornamentation. These specimens are assigned to *Acanthochitina* cf. *cancellata* Martin, 1983 (Fig. 5A–G). The second most common genus is *Spinachitina*, specimens of which are here assigned to *Spinachitina suecica* (Laufeld, 1967) (Fig. 5H–M). Because of poor preservation, several other chitinozoan specimens are kept under open nomenclature within the genera *Spinachitina* (Fig. 5N), *Conochitina* (Fig. 5O), *Lagenochitina?* (Fig. 5P), and *Tanuchitina* (Fig. 5Q–R).

## Discussion

The regional fossil record and facies of the Takche Formation indicate shallow-water conditions within the subtropical–tropical realm (Bhargava and Bassi 1986, 1998; Bordoloi 2002; Suttner 2003; Suttner et al. 2007b; Chaubey et al. 2023) that usually offer ideal conditions for jawed polychaetes (Eriksson et al. 2004). The two most productive samples were P/8/69 and P/13/231, both of which are on top of a transgressive cycle (sensu Suttner et al. 2007b). The preservation of maxillae allowed recognition of a general pattern only, which shows that, e.g., the labidognath taxa prevail compared to placognaths. As computed tomography (CT) has become a major method in palaeontology for non-destructive analysis and 3D reconstruction of fossil invertebrate and vertebrate soft and hard tissue (Abel et al. 2012), we decided to apply the method to the specimens herein.

There is only a handful of papers on CT studies of scolecodonts. Clemo and Dorgan (2017) undertook an analysis of Recent jawed polychaetes, mainly focusing on the maxillary apparatuses. The resulting 3D models are based on computed tomography data produced for analysis of the principles of the functionality of the jaw apparatuses. Recently, Eriksson et al. (2017) analysed fossil jaws using CT scanning for the reconstruction of maxillae that were embedded in sedimentary rock. They studied the oldest bobbit worm from the Lower–Middle Devonian of Ontario, Canada. However, the maxillae were over one centimetre in length, which is extreme among scolecodonts. Parry et al. (2019) employed micro-CT to reconstruct the maxillary apparatus of *Plumulites* (machaeridian) and confirmed a long-dis-

puted position of machaeridians within the crown group of the Phyllocodida (sister group of Eunicida). Thanks to micro-CT analysis, Gueriau et al. (2023) resolved a genuine affinity of *Gilsonicaris rhenanus* from the Lower Devonian Hunsrück Slate, Germany and assigned the specimen to eunicidan polychaetes.

We have chosen the new genus and species *Spitiprion khannai* Tonarová, Suttner, & Hints to be documented using micro-CT and submicron-CT, respectively. It is also the most characteristic species among the entire scolecodont assemblage obtained. Its maxillae are relatively large and sturdy, even though their length varies around 500 µm. For the isolated maxillae, the resolution of micro-CT turned out to be too low; therefore the submicron-CT was applied (parameters in Table 1; results in videos SOM 1–6). The micro-CT was applied for the partly preserved apparatus, embedded in the piece of rock (sample P/8/69, Fig. 4). The CT analysis allowed us not only to document the outer surface of the maxillae but also to study the myocoele cavity and observe tiny cavities in the maxillae walls. The polygonal internal structure is also visible.

In comparison to other regions, the diversity of ramphoprionids is much lower, with just one genus present. Elsewhere, the family is represented by the genera *Ramphoprion* Kielan-Jaworowska, 1962, and *Protarabellites* Stauffer, 1933 (see, e.g., Hints 2000). However, to date all Late Ordovician Gondwanan assemblages studied bear specific endemic taxa among ramphoprionids. In Spiti, it is *Spitiprion khannai* Tonarová, Suttner, & Hints gen. et sp. nov., in Saudi Arabia “Gen. et sp. indet. 1” of Hints et al. (2015) and *Ramphoprion* sp. according to Tonarová et al. (2023) in the Prague Basin. Even though, the last species is rather similar to the Laurentian species *Ramphoprion* cf. *deflexus*.

Except for the endemic *Spitiprion*, the Indian assemblage shares a general pattern with other localities from Gondwana. There are recorded polychaetaspids, a very common family in Ordovician and Silurian samples worldwide (Eriksson et al. 2013). Even though the diversity of at least two species of the genus *Oeononites* Hinde, 1879, is lower than observed in coeval strata (Hints 2000; Tonarová et al. 2023). On the Gondwanan shelf, paulinitids of the genus *Kettnerites* are first known from the uppermost Katian of Saudi Arabia (Hints et al. 2015), and in the lowermost Hirnantian in the Prague Basin (Tonarová et al. 2023), which is corroborated by the occurrence of *Kettnerites* sp. in the sample P/8/69. Placognaths, represented by *Vistulella* and *Mochtyella*, are relatively rare in the Pin River section. Placognath mochtyellids were also described from the Hirnantian Soom Shale Lagerstätte in South Africa by Whittle et al. (2008), who raised a new genus and species *Synaptogenys rietvleiensis*. On the other hand, an otherwise common genus in the Ordovician of Baltica, *Pistoprion* Kielan-Jaworowska, 1966, is missing in Indian samples, similarly as in the Prague Basin (Tonarová et al. 2023), where it first appeared during the Llandovery (Tonarová et al. 2019). Xanioprionids are very rare in Spiti. However,

Whittle et al. (2008) described *Xanioprion?* n. sp. from the Hirnantian Soom Shale Lagerstätte in South Africa, and the family is a common component of Late Ordovician assemblages in Baltica (Hints 2000; Hints et al. 2010).

In summary, the generic composition is rather similar to that in other Gondwanan assemblages (Whittle et al. 2008; Hints et al. 2015; Tonarová et al. 2023), supporting the closer affinity of the Gondwanan assemblage to Laurentia. On the other hand, two families, Hadoprionidae and Kalloprionidae, that are present in Laurentia, have not been recorded in the Indian samples so far. Differences with Laurentia are also among the placognath and ctenognath taxa that were not present in the Upper Ordovician Sylvan Shale in Oklahoma, USA (Eriksson et al. 2005) but were found in the Upper Ordovician of the Cincinnati region (Eriksson and Bergman 2003) and abundantly on Baltica (Hints 2000).

The dark colour of chitinozoan vesicles supplementary to a regional conodont colour alteration index (CAI) of 4–5 (Suttner 2003) and 4 (Myrow et al. 2019), suggests a considerable thermal exposure (Tricker et al. 1992). That supports the suggestion of Sinha and Verniers (2016), who described also very dark vesicles of chitinozoans from the Kumaon region and interpreted the state of preservation as a result of intense tectonic activity during the Himalayan orogenic movement, which resulted in elevated burial temperatures.

Apart from the contribution to the regional thermal history, chitinozoans are useful climate markers, as shown e.g., by Vandenbroucke et al. (2010), who reconstructed palaeoclimatic belts based on the palaeogeographic distribution of Late Ordovician chitinozoan assemblages. At the Pin River section, five chitinozoan genera were identified and compared regionally to the findings from the closely located Parahio River section by Wang et al. (2021). Our results show a less diverse assemblage than that of Wang et al. (2021), lacking the genera *Euconochitina*, *Cyathochitina*, and *Ancyrochitina*. Another two genera missing in the Pin River assemblage are *Angochitina* and *Belonechitina*, which were described from the Shiala Formation in Kumaon by Sinha et al. (2011) and Sinha and Verniers (2016). On the other hand, representatives of *Conochitina* were documented from both Spiti and Kumaon.

Specimens of *Acanthochitina* cf. *cancellata* most closely resemble *Acanthochitina? cancellata* of Martin (1983) from lower Katian of Quebec in test outline and size. However, the type specimen of *Acanthochitina? cancellata*, together with the subsequent reports from Quebec (Achab 1978; Asselin et al. 2004), is photographed using an optical microscope; thus, fine morphological details and the ornamentation of this species have remained poorly known.

The second common species is *Spinachitina suecica*, which, according to previous publications, is mainly reported from the Sandbian to lower Katian. The type specimen comes from the middle Sandbian upper Dalby Limestone of Sweden (Laufeld 1967). Later, *S. suecica* was reported from the *Macrourus* siltstone from the Öland erratics (middle Caradocian according to Grahn 1981), the upper Kukruse

to Nabala Baltic regional stages (lower and middle Katian) of Gotland, Sweden (Grahn 1982), the uppermost Uhaku–middle Haljala regional stages (uppermost Darriwilian–mid Sandbian; Middle–Upper Ordovician) of Estonia (Nölvak and Bauert 2006). The material recovered in the present study shows some morphological variability, from a typical shape of the holotype (Fig. 5H) to a more slender form (Fig. 5M). The slender form also resembles *Spinachitina* sp. aff. *coronata* from the *Ancyrochitina merga* biozone (middle upper Katian) of Central Anti-Atlas, Morocco (Bourahrouh et al. 2004). However, the type material of *Spinachitina coronata* recovered in Baltica is characterised by a long and slender test of around 300 microns and a poorly developed flexure. Generally, the chitinozoan assemblage from the Takche Formation indicates a Late Ordovician age.

## Conclusions

- Scolecodonts and chitinozoans from the Upper Ordovician Pin Member of the Takche Formation (Katian, *Amorphognathus ordovicicus* Conodont Biozone) at the Pin River section, Pin Valley, Spiti, northern India, are reported.
- A collection of approx. 60 scolecodonts, assigned to *Oeononites*, *Atraktoprion*, *Vistulella*, *Kettnerites*, *Mochtyella*, *Xanioprion*, and the new genus *Spitiprion*, and approx. 50 chitinozoans belonging to *Acanthochitina*, *Conochitina*, *Lagenochitina?*, *Spinachitina*, and *Tanuchitina* were recovered.
- A new ramphoprionid genus and species, *Spitiprion khananai* Tonarová, Suttner, & Hints is described based on a partly preserved maxillary cluster and isolated maxillae. To date, it is endemic to the Himalayan region.
- Comparison with other Gondwanan jawed polychaete faunas, e.g., from the Hirnantian Soom Shale in South Africa, indicates that both areas, even though located on opposite sides of the Gondwanan shelf, have representatives belonging to Mochtyellidae and Xanioprionidae in common. Geographically closer assemblages from Saudi Arabia share besides placognath jaws even more taxa at the genus level including *Atraktoprion*, *Kettnerites*, and *Oeononites*.
- This study is the first to present submicron-CT data on scolecodonts. We have confirmed the usefulness of this technique on further jawed polychaete studies, contributing to the establishment of 3D virtual scolecodont type collections.

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## Authors' contributions

Conceptualisation, funding acquisition, and project administration PT; resources and data curation PT, TJS; formal analysis PT, OH, YL; investigation PT, OH, TJS, YL; methodology and software PT, MZ, MK, TZ, JK; visualization PT, MZ, MK, TZ, JK, TJS; writing: original draft, review and editing PT, TJS, OH, MZ, MK, YL, TZ, JK, EK.

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